

Review Article

Indian J Med Res 132, August 2010, pp 129-149

Cancer biomarkers - Current perspectives

Anant Narayan Bhatt, Rohit Mathur, Abdullah Farooque, Amit Verma & B.S. Dwarakanath

Division of Radiation Biosciences, Institute of Nuclear Medicine & Allied Sciences, Delhi, India

Received November 21, 2008

In the recent years, knowledge about cancer biomarkers has increased tremendously providing great opportunities for improving the management of cancer patients by enhancing the efficiency of detection and efficacy of treatment. Recent technological advancement has enabled the examination of many potential biomarkers and renewed interest in developing new biomarkers. Biomarkers of cancer could include a broad range of biochemical entities, such as nucleic acids, proteins, sugars, lipids, and small metabolites, cytogenetic and cytokinetic parameters as well as whole tumour cells found in the body fluid. A comprehensive understanding of the relevance of each biomarker will be very important not only for diagnosing the disease reliably, but also help in the choice of multiple therapeutic alternatives currently available that is likely to benefit the patients. This review provides a brief account on various biomarkers for diagnosis, prognosis and therapeutic purposes, which include markers already in clinical practice as well as various upcoming biomarkers.

Key words Biomarkers - cancer - cancer therapy - diagnosis - glycolysis - molecular targets - Pin 1 - prognosis

Introduction

Every cell type has a unique molecular signature, referred to as biomarkers, which are identifiable characteristics such as levels or activities (the abilities of genes or proteins to perform their functions) of a myriad of genes, proteins or other molecular features. Biomarkers are therefore, an objective measure or evaluation of normal biological processes, pathogenic processes, or pharmacological responses to a therapeutic intervention¹. This includes all diagnostic tests, imaging technologies, and any other objective measures of a person's health status. Biomarkers are subject to dynamic modulation, and are expected to enhance our understanding of drug metabolism, drug action, efficacy, and safety. These can also facilitate molecular definition of diseases, provide information

about the course of disease and predict response to therapies.

More than 11 million people are diagnosed with cancer every year. It is estimated that there will be 16 million new cases every year by 2020². Cancer is a cluster of diseases involving alterations in the status and expression of multiple genes that confer a survival advantage and undiminished proliferative potential to somatic or germinal cells³. Alterations primarily in three main classes of genes *viz.*, (proto) oncogenes, tumour suppressor genes and DNA repair genes collectively contribute to the development of cancer genotype and phenotype that resists the natural and inherent death mechanism(s) embedded in cells (apoptosis and like processes), coupled with dysregulation of cell proliferation events (Fig.). There is increasing

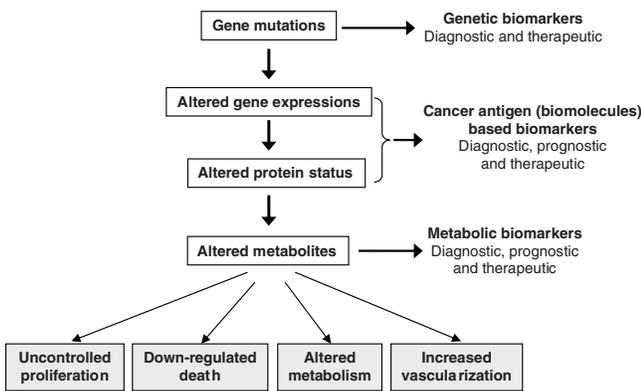


Fig. The process of carcinogenesis, showing opportunities of identifying biomarkers.

evidence to suggest that cancer is also driven by 'epigenetic changes' like DNA methylation and altered patterns of histone modifications, leading to alterations in chromatin condensation status thereby regulating expression of certain set of specific genes^{4,5}.

Technologies to recognize and understand the signatures of normal cells and how these become cancerous, promises to provide important insights into the aetiology of cancer that can be useful for early detection, diagnosis, and treatment. Biomarkers are therefore invaluable tools for cancer detection, diagnosis, patient prognosis and treatment selection⁶. These can also be used to localize the tumour and determine its stage, subtype, and response to therapy. Identification of such signature in surrounding cells or at more distal and easily sampled sites of the body *viz.*, cells in the mouth (instead of lung) or urine (instead of urinary tract) can also influence the management of cancer.

Cancer cells display a broad spectrum of genetic alterations that include gene rearrangements, point mutations, and gene amplifications, leading to disturbances in molecular pathways regulating cell growth, survival, and metastasis. When such changes manifest in majority of patients with a specific type of tumour, these can be used as biomarkers for detection and developing targeted therapies, besides predicting responses to various treatments⁷⁻⁹.

Genetics, genomics, proteomics, many non invasive imaging techniques and other technologies allow measurement of several biomarkers. Currently, there is a greater understanding of the disease pathways, the protein targets and the pharmacologic consequences of drug administration. Therefore, application of

biomarkers in the clinical practice is likely to result in advanced knowledge leading to a better understanding of the disease process that will facilitate development of more effective and disease specific drugs with minimal undesired systemic toxicity. Establishment of biomarkers requires a comprehensive understanding of the molecular mechanisms and cellular processes underlying the initiation of cancer, especially focusing on how small changes in only a few regulatory genes or proteins can disrupt a variety of cellular functions. A major challenge in cancer diagnosis is to establish the exact relationship between cancer biomarkers and the clinical pathology, as well as, to be able to non invasively detect tumours at an early stage. Similarly, identification of subtle changes in the genomics and proteomic status specific to malignant transformation will allow molecular targets to be used for developing therapeutics. This review is a brief account of the biomarkers employed currently in clinical oncology for diagnosis and therapy as well as potential ones that particularly hold promise as targets for therapy.

Diagnostic and prognostic biomarkers are quantifiable traits that help clinical oncologists at the first interaction with the suspected patients. These particularly aid in (i) identifying who is at risk, (ii) diagnose at an early stage, (iii) select the best treatment modality, and (iv) monitor response to treatment⁶. These biomarkers exist in many different forms; traditional biomarkers include those that can be assessed with radiological techniques *viz.*, mammograms *etc.*, and circulating levels of tumour specific (related) antigens for example, prostate-specific antigen (PSA). With the availability of complete human genome sequence, and advancement in key technologies such as high-throughput DNA sequencing, microarrays, and mass spectrometry, the plethora of potentially informative cancer biomarkers has expanded dramatically to include the sequence and expression levels of DNA, RNA, and protein as well as metabolites¹⁰. Advances in imaging technologies open up the possibility that pertinent molecular biomarkers (*e.g.*, those marking response to therapy) can be monitored in cancer patients non invasively.

Genetic and genome based approaches have played significant role in the diagnosis and prognosis of cancer. Alterations in the DNA content (hyper- and hypo- diploidy) arising on account of genomic instability during dysregulated proliferation has been extensively used, but has its limitations. Structural anomalies in chromosomes on the other hand are not a

universal phenomenon. Molecular genetic techniques providing information about the specific and subtle genetic changes have been quite useful in the identification of certain tumours. With the introduction of several comparative genetic analysis techniques, the detection of tumours by analyzing subtle changes in the genetic composition has become more feasible. Analysis of global gene expression changes provided by the micro-array technology has revolutionized the genome based approaches for studying biomarkers at large and cancer in particular. These techniques have in fact been quite successful in clearly dissecting subtle changes between different stages of tumours as well as resolving similar tumour types, which were otherwise not easily discernable.

Cytogenetic and cytokinetic markers

Structural and numerical aberrations in the chromosomes are classical markers of cancer as the association between chromosomal aberrations and neoplastic transformation has been well established. While deviations from diploid chromosome number leading both to hyper- and hypo-diploidy as well as aneuploidy have been noted in malignant tumours¹¹, sister chromatid exchanges and translocations give rise to structural aberrations that can be easily scored using various banding techniques. Further, double minutes and homogeneously stained regions (indicative of gene amplification) are also often observed in malignant cells that can serve as markers¹². Although, the ploidy changes complement the clinico-pathological findings, a weak association between ploidy, histological and clinical staging has been noted in many tumours¹³. Somatic mutations (in reporter genes, oncogenes and tumour suppressor genes) are promising biomarkers for cancer risk as these can capture genetic events that are associated with malignant transformation¹⁴. There is growing evidence that specific polymorphism in certain genes are associated with cancer risk^{15,16}.

Among other genome based biomarkers, identification of neoplasm from the level of lesion specific transcriptomes (mRNA of cytokeratin-19, EGFR, MUC 1, *etc.*) in the blood has been successfully employed in certain epithelial tumours^{17,18}. Recently a novel transcriptome marker based on the levels of exon-3 deleted variant isoform of proghrelin (ghrelin is a growth factor involved in prostate cancer cell proliferation) has been developed that aims at reducing the false positives in prostate and other endocrine cancers¹⁹.

Enhanced cell proliferation is one of the most important hallmarks of cancer, which is easy to identify using a number of histological, biochemical and flow cytometric analysis. Although subjective in nature, histological assessment based on evaluating the number of mitotic cells present within a given sample, is still used as a routine clinical test and even for grading in certain tumours like breast cancer. Flow cytometric analysis of DNA content, which is automated, objective and rapid allowing large number of cells (and samples) to be measured, has been extensively used for the assessment of proliferation status. This complements histological analysis in most cases, besides allowing analysis of clonal and spatial heterogeneity, two important hallmarks of highly malignant tumours¹¹. Identification of S-phase cells (unequivocal marker of proliferation) and analysis of a number of other antigenic determinants of proliferation (PCNA, Ki67 NOR, *etc.*) studied using a variety of cell biology techniques have also been used as complementary markers. Information provided by gene expression analysis has a distinct advantage over other assessments of proliferation (*viz.*, more quantitative, objective, and automated) and could form a component of genomic-based clinical diagnostics of cancer. Proteins encoded by the minichromosome maintenance (*MCM*) genes have also been proposed as useful markers of proliferation; with high levels of gene expression indicating poor prognosis²⁰. All these genes are cell-cycle regulated and are found among the genes associated with proliferation in tumors²¹.

Genetic biomarkers

Cancer is a genetic disease initiated by alterations in genes, such as oncogenes and tumour suppressors that regulate cell proliferation, survival, and other homeostatic functions. Gain/ loss of gene function is predominantly responsible for oncogenic transformation. Several proto-oncogenes get converted into oncogenes with as little as a point mutation on a chromosome, thereby altering the amount of its product *i.e.*, protein. Several non-random mutations, and translocations/ rearrangement within the regulatory region of the gene are also known to be associated with particular types of malignancy. For example, the "Philadelphia chromosome" is associated with chronic myelogenous leukaemia due to a translocation between chromosomes 9 and 22. Other examples occur in Burkitt's lymphoma and in follicular B-cell lymphomas. These translocations serve as highly specific tumour markers for unique clinical diagnosis.

Extensive allotyping of breast cancer for gene deletions of loci on multiple chromosomes has been reported²². Deletion of genomic material is important, because the lost segment of DNA may contain certain tumour suppressor activity. Gene deletions are discovered by polymerase chain reaction (PCR) using microsatellite probes to various chromosomes and sites. Tumour-suppressor genes are thought to play a role in specific tumour *e.g.*, in breast tumour p53, Rb, DCC, Brush-1, BRCA-1, BRCA-2. In addition, allelic losses with more or less significant breast carcinoma associations on virtually all chromosomes have been reported²³. Unfortunately, unlike these well defined markers, random chromosomal abnormalities that are not associated with a particular morphological change give rise to clinical cancer²⁴.

Loss of heterozygosity as well as mutations within several protooncogenes can lead to microsatellite instability (MSI)²⁵. Although detection of microsatellite instability/alterations in pathologic tissue samples require a comparison with normal tissue but it presents a valuable tool for early detection, occasionally at preneoplastic stage²⁶. MSI can also be used for prognosis and evaluation for chemotherapeutic response²⁵.

Adenomatous polyposis coli (APC) gene: The *APC* gene normally responsible for suppressing cancer, is deactivated in many tumours, the altered gene has been found in 92 per cent of patients diagnosed with oesophageal adenocarcinoma, and in 50 per cent of patients with squamous cell carcinoma of the oesophagus²⁷. Mutations in the *APC* gene occur in 60 per cent of patients with colorectal carcinoma and are thought to be the earliest genetic abnormalities in the progression of colorectal carcinoma²⁸. Most of these mutations cause the production of an *APC* protein that is abnormally short and nonfunctional. This short protein cannot suppress the cellular overgrowth that leads to the formation of polyps, which could become cancerous²⁸. Somatic mutation of the *APC* gene has been identified in sporadic colorectal cancer as well as in some cancer of the stomach, pancreas, thyroid, ovary and other primary sites²⁸. Hence, hypermethylated *APC* gene is being utilized as a biomarker to determine the stage of oesophageal cancer, detect recurrent disease, and monitor disease progression or treatment response²⁷. A high level of hypermethylated *APC* gene in the bloodstream is generally associated with poor survival; conversely, the prognosis improves dramatically for patients with low or undetectable blood levels of the altered gene²⁹. *APC* expression is also induced during

pregnancy and lactation in the mouse mammary gland, and *APC* deficiency results in defective lobule-alveolar development, therefore should be used with caution. PCR-based tests are widely used to detect these mutations.

Epigenetic biomarkers

In cancer cells, genes and their functional products are either modified by mutations, or through epigenetic modifications to chromosomes that alter gene-expression patterns. Epigenetic modifications can occur directly through DNA methylation of genes or indirectly by methylation, acetylation, or phosphorylation of histones and other proteins around which DNA is wound to form chromatin³⁰. DNA methylation at the cytosine residue is the main epigenetic modification in humans which occurs in the context of 5'-CpG-3' dinucleotides³¹. In recent years it has become apparent that epigenetic events are potentially responsible for cancer initiation and progression as genetic abnormalities, with DNA hypo- and hyper-methylation promoting cancer development. Several studies have shown that the activity and DNA methyltransferases (DNMTs), which add methyl groups to DNA at cytosine residues, are altered in tumour cells and associated with several developmental abnormalities³². Genomic hypomethylation may lead to both genomic instability and stronger gene expression³³. On the other hand, local promoter CpG island hypermethylation induces the functional silencing of tumour suppressor genes, mimicking their genetic mutations. Many genes involved in the process of carcinogenesis are the common targets for silencing, which includes cell-cycle control and apoptosis genes *viz.* *p14*, *p15*, *p16*, *Rb*, *DAPK*; DNA repair genes *MGMT*, *hMLH1*; adhesion and metastasis genes *CDH1*, *CDH13*; biotransformation genes *GSTP1* and signal transduction genes *RAR β* , *APC*³⁴. Since, methylation pattern of many genes are altered in cancer, type specific panels for methylation of different genes have been suggested, *e.g.*, *GSTP1*, *RAR β* , *TIG1* and *APC* for prostate carcinoma; *p16*, *RASSF1A*, *FHIT*, H-cadherin and *RAR β* for non small cell lung cancer³⁴; *VHL*, *p16*, *p14*, *APC*, *RASSF1A* and *Timp3* for kidney cancer, *etc*³⁴.

Hypermethylation markers may be used for the detection of both primary and metastatic or recurrent cancer cases. For example, hypermethylation of *p16* promoter in the circulating serum DNA correlate well with recurrent colorectal cancer³⁶. Aberrant methylation of the *p16Ink4* and *MGMT* promoters can be detected in DNA from the sputum of patients with squamous

cell carcinoma nearly 3 yr before clinical diagnosis while, methylation of *p16Ink4*, *RASSF1A*, or *PAX5-beta* genes appears to be associated with a 15-fold increase in the relative risk for lung cancer. Therefore, it has been suggested that alterations in methylation patterns of groups of genes in sputum samples may be an effective, non invasive approach for identifying smokers at risk of developing lung cancer³⁷. Methylation of the O6-methylguanine-DNA methyltransferase (*MGMT*) gene, which encodes a DNA-repair enzyme has been shown to inhibit the killing of tumour cells by alkylating agents and methylation of the *MGMT* promoter of malignant glioma appears to be a useful predictor of the responsiveness to alkylating agents as patients with silencing of this gene seem to respond better to therapy³⁸. Although, research in epigenetics has led to improved survival of patients with certain forms of lymphoma and leukaemias through the use of drugs that alter DNA methylation and histone acetylation, proposed methylation markers need further optimization and large scale clinical trial for further validation. The development of therapeutics that reverse epigenetic alterations in cancer cells, along with prognostic and diagnostic assays based on gene-methylation patterns, are promising new avenues for future improvements in patient care.

Cells as biomarker

In advanced stages of tumours, cells starts appearing in bloodstream where it can be easily monitored. Advanced clinical practice in certain malignancy have effectively used tumour and immune cells where it served as a good biomarker of prognosis, while its utility in other cancers are under evaluation at the present time.

Circulating tumour cells (CTCs): It is simple yet powerful biomarker in the field of oncology. The presence of CTCs has been shown to predict survival in patients with metastatic breast cancer at multiple time points throughout the course of therapy³⁹. CTCs provide an early, reliable indication of disease progression and survival for patients on systemic therapy for metastatic breast cancer. Elevated CTCs at any time during therapy is a harbinger of progression, while elimination of CTCs indicates effectiveness of the therapy⁴⁰. Selection of appropriate therapeutic regime can also be guided by assessment of the presence of the therapeutic target on the CTCs and, in contrast to CTC scans, the effect of therapy can be measured after the first cycle of therapy. CTCs are not only potential

surrogate endpoint in oncology clinical trials but may guide the selection of patients into the trials⁴¹.

CTCs have been shown to be superior to standard tumour markers (*e.g.*, Ca27-29) in predicting prognosis. Furthermore, the efficacy or benefit to systemic therapy can also be predicted by the level of CTCs as early as 3-4 wk after initiation of therapy. Patients with persistent CTCs (≥ 5) demonstrate lack of response to treatment or progressive disease at the time of restaging by standard imaging modalities, while objective remission have been reported in patients with < 5 CTCs. Available evidences clearly suggest that CTCs can be used as an early predictor of treatment efficacy and extremely useful in sparing patients from futile therapy early in the course of their treatment⁴².

T-regulatory cells (CD4⁺, CD25⁺ and Foxp3⁺): A number of mechanisms contribute to the capacity of the immune system to discriminate self from non-self, facilitating the maintenance of immunological tolerance to self-antigens and the induction of protective immunity to foreign antigens. It is becoming increasingly clear that regulatory T cells (T-regs) are equally important in inducing and maintaining peripheral self-tolerance and thus preventing immune pathologies^{43,44}. These are subpopulations of CD4 cells, characterized by high CD25 expression along with Fox P3. They are thought to play a specialized role in controlling both innate and acquired immune responses⁴⁵. Furthermore, studies in cancer patients suggests that increased T-regs activity may be associated with poor immune responses to tumour antigens and contribute to immune dysfunction resulting in tumour growth^{46,47}. High numbers of T-regs have been found in lung, pancreatic, breast, liver and skin cancer patients, either in the blood or in the tumour itself^{46,48-51}. However, levels of T-regs are also elevated in certain infectious diseases⁵². T-regs are able to inhibit proliferation and IFN- γ production by CD4⁺ and CD8⁺ T cells, as well as natural killer (NK) cell-mediated cytotoxicity. Studies on ovarian carcinoma patients have demonstrated that the presence of T-regs, which suppress tumour-specific T-cell immunity inversely correlate with survival⁵³. Recent studies in Ehrlich ascites tumour bearing mice have also shown that higher numbers of T-regulatory cells are associated with tumour progression. Further, the T-regs levels were found to be significantly lower in animals that showed complete response (tumour regression and cure) to a given treatment (a combination of radiation and the glycolytic inhibitor 2-deoxy-D-glucose;2-DG), while an increase was found in those animals that did

not respond to the treatment suggesting a relationship between T regulatory cells and therapeutic response⁵⁴. Therefore, it appears that although T-regs may serve as a surrogate immune marker of cancer progression (and perhaps prognosis), it seems to be more useful as a predictor of response to therapies.

Regulatory T cells (T-regs) are primarily and specifically identified by the experiments of (transcription factor, FoxP3) using anti FoxP3 antibodies. The presence of FoxP3⁺ cells within tumours has been shown to predict the prognosis, invasiveness, and metastatic ability of some tumours by modulating the ability of the immune system to target tumour cells⁵⁵. Depletion of regulatory T cells from tumours could lead to the rejection of both early- and late-stage tumours by the host immune system. These findings suggest that the widespread use of T-regs, and more importantly its intracellular marker FoxP3, should be explored as a biomarker for human tumours to enabling better decisions in patient care, as well as prepare the field for novel therapeutic agents directed at the elimination of regulatory T cells within tumours or in the peripheral blood.

Cancer stem cells (CSCs): It has long been recognized that subpopulations of cancer cells exist within the tumours that resemble the developmental hierarchy of the normal tissue from which the tumour arose. In recent years, the cancer stem cell model of tumourigenesis has received increasing attention. This model postulates that tumours are driven and maintained by a minority subpopulation of cells that have the capacity to self-renew and to generate the more differentiated progeny which make up the bulk of a tumour⁵⁶. The former population has been termed cancer stem cells (CSCs), tumourigenic cancer cells, or tumour-initiating cells, by various investigators, to indicate that only these can give rise to new tumours when transplanted into immuno-deficient animals⁵⁷.

Evidence for the existence of CSCs initially came from studies of acute myelogenous leukemia (AML). Presence of CSCs have now been demonstrated in many solid tumours, including glioblastoma, medulloblastoma, breast cancer, melanoma, and prostate cancer⁵⁸. The existence of CSCs has profound implications for cancer biology and therapy because it is likely that eradication of CSCs is the critical determinant in achieving cure. It has been proposed that CSCs may be particularly resistant to chemotherapy and radiation therapy as has been shown in a study with glioblastoma⁵⁹. CD133⁺ cells were earlier suggested as

the tumourigenic population in primary glioblastoma multiforme specimens⁶⁰; while more recent studies have shown that these are indeed more radioresistant compared with CD133⁻ tumour cells, as their fraction increase after irradiation which appears to be mainly responsible for the tumour regrowth⁵⁹. Therefore, it appears that identifying and characterizing CSCs for every possible tumour is of paramount importance and will likely lead to new therapeutic avenues. Also, work on radiosensitizers should begin to focus on preferentially affecting CSCs compared with normal tissues and normal tissue stem cells.

Viral biomarkers

Among viral induced cancers, hepatocellular carcinoma (HCC) is one of the most common cancers worldwide and a leading cause of death in developing countries, where nearly 80 per cent of the cases are reported⁶¹. Risk factors include chronic hepatitis infections mainly due to the endemic hepatitis B virus (HBV) infection, whereas association of hepatitis C virus (HCV) infection is also reported in a small fraction (12 - 17%) of the HCC cases⁶¹. Beside immunoinflammatory reactions, HBV can also promote carcinogenesis through genetic instability generated by its common integration in host DNA⁶². A number of different types of biomarkers have been used to understand the aetiology and progression of HCC. Perhaps, the most well known are the serum/plasma markers of HBV or HCV infection⁶². These markers include analysis of viral DNA or proteins or antibodies produced against the viral proteins. HBV surface antigen (HBsAg) is most frequently used to determine chronic infection with high or low viral replication, while HBeAg is a measure of chronic infection with high viral replication⁶³. The other major classes of biomarkers used in studies of HCC are analysis of antibodies including measurement of anti-HBV core antigen, anti-HBe antigen and anti-HBsAg^{61,63}.

Cervical cancer is the second most common cancer and predominant gynaecological cancer in women, causing most cancer related deaths world over⁶⁴. There are several factors which contribute to high incidence of this disease are early age of marriage, multiple sexual partners, multiple pregnancies, poor genital hygiene, smoking and use of oral contraceptives. But the most predominant aetiological factor for cervical cancer is persistent infection of certain high-risk types of human papillomaviruses (HR-HPVs), while low risk types are associated with benign cervical lesions and genital warts⁶⁵. HPV has also been detected in a significant

proportion of oral, oesophageal, anal, vaginal, vulvar, and penile cancer and in a small percentage of lung, laryngeal, and stomach cancer, as has been shown in some parts of the world⁶⁶. HPV viral load, a measure of the amount of viral DNA in biopsy specimens, alone or in conjunction with well characterized HPV serologic assays, has been suggested to delineate the role of HPV among oral and oropharyngeal cases⁶⁶, while antibodies generated in the subjects against HPV E6 and E7 serve as markers of an invasive HPV-associated malignancy⁶⁷. Viral load assessment can also be exploited to distinguish clinically relevant HPV infections in the cervix⁶⁴. Prophylactic immunization of women who are negative for the HPV16 L1, E6 and E7 oncoprotein markers, has been reported to eliminate their risk for HPV16-related cervical intraepithelial neoplasia⁶⁷. Recently, the two new HPV vaccines “Gardasil” and “Cervarix” have been shown to be highly immunogenic and effective in preventing infection with high-risk HPV types 16 and 18, the two most common oncogenic types associated with this disease⁶⁵. Papillomaviruses were first identified, cloned and sequenced from cervical tumour specimens and subsequently established as important causative agents for development of cervical cancer⁶⁶.

Epstein-Barr virus (EBV) was the first human virus to be directly implicated in carcinogenesis. It infects more than 90 per cent of the world’s population, out of which a small proportion develop tumours⁶⁸. Although herpesviruses are ubiquitous in nature, interestingly humans serve as the only natural host for EBV and upon infection; the individual remains a lifelong carrier of the virus⁶⁸. The vast majority of the world’s population exhibits antibodies to EBV and the infection usually occurs early in childhood⁶⁸. EBV has been implicated in the pathogenesis of Burkitt’s lymphoma, Hodgkin’s disease, non-Hodgkin’s lymphoma, nasopharyngeal carcinoma, and lymphomas, as well as leiomyosarcomas arising in immuno-compromised individuals⁶⁸. EBV infection of B lymphocytes is thought to occur in the oropharyngeal lymphoid organs, and in normal carriers, while the virus persists in circulating memory B cells. EBV is a herpesvirus with a 184-kbp long, double-stranded DNA genome that encodes more than 85 genes⁶⁸. The EBV genome is maintained in B cells, either as a multicopy circular episome in the host cell or by integrating the viral DNA into the host genome. The virus thus ensures transmission to cell progeny when B lymphocytes replicate. EBV DNA in the peripheral blood predicted a high risk of distant

metastases⁶⁹. Detection and quantification of plasma EBV DNA serves as a useful molecular marker for diagnosis, monitoring, and prediction of relapse in patients with nasopharyngeal carcinoma and Hodgkin’s lymphoma^{69,70}.

Thus viral biomarkers seem to have potential application in the diagnosis (particularly staging), prognosis and predicting as well as monitoring response to therapy.

Cancer antigens (biomolecules) based biomarkers

The cancer proteome contains information on perhaps every biological process that takes place in cancer cells, cancer tissue microenvironment, and cancer cell-host interaction. Cancer cells release many proteins and other macromolecules into the extracellular fluid through secretion that can also serve as biomarkers. Some of these products can end up in the bloodstream and hence serve as potential serum biomarkers. Some important cancer antigens that serve as diagnostic and prognostic biomarkers of cancer are summarized in the Table.

Prostate specific antigen (PSA), a 33 kDa serine protease belonging to the family of “Kallikrein genes” and produced by both normal as well as neoplastic prostate epithelial cells is the most widely studied biomarker in prostate cancer. Among all kallikreins, hK2 and hK3 expression is highly restricted to the prostate in males and are therefore useful as biomarkers⁷¹. Being a protease, it appears to be involved in the initiation and growth of prostate cancer by abnormal release of growth factors or proteolysis of growth factor binding proteins. It may also have a role in invasion and metastases through the degradation of collagen and laminin. PSA was first identified by in 1971 while attempting to find a substance in seminal fluid that would aid in the investigation of forensic cases. PSA was first measured quantitatively in the blood by Papsidero and colleagues in 1980, who also reported its clinical use as a marker of prostate cancer⁷². PSA is present in small quantities in the serum of normal men, and is often elevated in the presence of prostate cancer and other prostate disorders. However, prostate cancer can also be present in the complete absence of an elevated PSA level⁷³. PSA expression is androgen dependent and therefore less sensitive in older population. Obesity has been reported to reduce serum PSA levels⁷⁴, while increase has been found in prostate infection, irritation, benign prostatic hyperplasia (BPH), and ejaculation⁷⁵. Limitations of PSA as a biomarker for monitoring

Table. Cancer biomarkers for diagnosis and prognosis of the disease

Biomarker	Tumour	Application	Sample type/ Method of detection
<i>Cancer antigen (biomolecules) based biomarkers:</i>			
Prostate specific antigen (PSA)	Prostate cancer	Diagnostic and prognostic	Serum/ Immunoassay
Alpha-foetoprotein (AFP)	Hepatocellular carcinomas (HCC)	Diagnostic and prognostic	Serum/ Immunoassay
Cancer antigen 125 (CA125)	Ovarian cancers Fallopian tube cancer	Diagnostic and prognostic	Serum/ Immunoassay
Cancer antigen 15-3 (CA15-3)	Breast cancer	Diagnostic and prognostic	Serum/ ELISA, Lymph node/ IHC, Bone marrow/ IHC
Cancer antigen 19-9 (CA 19-9)	Pancreatic cancer Bladder cancer	Diagnostic and prognostic	Serum/ ELISA Urine/ ELISA
BRCA-1, BRCA-2	Breast cancer	Diagnostic	Tumour samples/ RT-PCR
Carcinoembryonic antigen (CEA)	Colorectal cancer	Diagnostic and prognostic	Serum/ ELISA
Human chorionic gonadotrophin (hCG)	Germ cell tumours (ovarian and testicular)	Diagnostic	Serum/ ELISA
Thyroglobulin (Tg)	Papillary and follicular thyroid cancer	Diagnostic and prognostic	Serum/ ELISA or IHC with TPO Ab
Heat shock proteins (HSPs) Hsp27; Hsp70	Gastric, prostate carcinoma, osteosarcomas, uterine, cervical, and bladder carcinoma	Diagnostic and prognostic	Serum/ ELISA
TGF β	Malignant tumours	Diagnostic and prognostic	Serum / ELISA
<i>Metabolic biomarker:</i>			
Glucose metabolism	All cancers, general	Daignostic, prognostic and therapeutic	Imaging/ FDG-PET scan
<i>Genetic biomarkers:</i>			
Genetic translocations viz. Philadelphia chromosome, Bcl2 and other gene translocation fusion products	AML, ALL, CML, MDS and Burkitt's lymphoma	Diagnostic	Bone marrow or peripheral blood/ FISH
APC gene	Adenocarcinoma, squamous cell carcinoma of the stomach, pancreas, thyroid and ovary	Diagnostic and prognostic	Blood, Tumour sample/ RFLP of chromosome 5q21-22, Methylation status of APC gene
<i>Cells as biomarker:</i>			
Circulating tumour cells (CTCs)	Metastatic breast cancer, etc.	Diagnostic and prognostic	Blood/ Immunocytometry
Cancer stem cells (CSCs)	AML, melanoma, brain tumour, breast cancer, prostate cancer	Diagnostic, prognostic and therapeutic	Tumour sample/ Immunocytometry

response to the therapy have been identified, as increase in serum level not correlating with tumour regression following radiotherapy has been reported in some instances⁷⁶. A blood test to measure PSA is considered the most effective test currently available for the early detection of prostate cancer.

Alpha-foetoprotein (AFP) is the major serum foetal protein in mammals, which is actively produced and secreted during the foetal life by the liver hepatocyte.

Major tumours that secrete AFP are endodermal sinus tumour (yolk sac carcinoma), neuroblastoma, hepatoblastoma, and hepatocellular carcinoma⁷⁷. It is a well-known diagnostic biomarker used to follow the development of hepatocellular carcinomas (HCC) where its synthesis is frequently upregulated. Significant increase in the amount of serum AFP is usually detectable in patients with poorly differentiated and highly malignant tumours⁷⁷. However, AFP test is

not sensitive or specific enough for early detection of the hepatocellular carcinoma and therefore by itself is not diagnostic, but could only be suggestive and useful as a prognostic marker. Since the levels of AFP may be elevated in serum from patients with other chronic liver disease; AFP is not useful for screening in patients suffering from cirrhosis or hepatitis C^{78,79}.

CA125: The CA 125 antigen is a membrane glycoprotein produced by tissues derived from coelomic epithelium that is expressed by most epithelial ovarian cancers⁸⁰. CA125 was initially detected in 1983 using the monoclonal antibody designated OC125; hence the name CA125. CA125 is a powerful index of risk of ovarian and fallopian tube cancer in asymptomatic postmenopausal women. It is found in the serum of more than 80 per cent of the patients with epithelial ovarian tumours, with half life of 4 days. CA-125 antigen remains the only serum tumour marker routinely used in epithelial cancer of the ovary for patient prognosis, disease progression, and response to chemotherapy^{80,81}. Normal values of CA125 in serum range from 0 to 35 U/ml. CA125 is also expressed by a number of tissues of both cancerous and noncancerous origin⁸². It may also be elevated in other malignant cancers, including those originating in the endometrium, fallopian tubes, lungs, breast and gastrointestinal tract⁸³. Certain physiological conditions also modulate CA125 levels, as the levels are elevated slightly during menstruation⁸⁴ and more prominently during the first trimester of pregnancy. A decrease in CA125 level is generally associated with tumour response to therapy, whereas a rising level is suggestive of drug resistance⁸⁵. Indeed, CA125 is an accurate marker to define relapse of ovarian cancer. CA125 has numerous applications in the design of clinical trials, from prognosis to follow up and is a tool that is complementary to standard criteria for disease measurement⁸⁶. The key problems in using the CA125 test as a screening tool are its lack of sensitivity and its inability to detect early stage cancers.

CA15-3: The CA15-3 protein is a member of the family of proteins known as mucins, whose normal function is cell protection and lubrication. It plays a role in reducing cell adhesion and is found throughout the body. Elevated levels of this antigen are found mainly in breast cancer where it appears to be involved in metastasis⁸⁷. CA15-3 level is elevated in nearly 11 per cent of women with operable breast cancer, and 60 per cent of women with metastatic disease. Pre-operative concentration of CA15-3 is associated with worse prognosis than those with low concentrations⁸⁸.

CA15-3 appears to be a marker for individualizing therapy in patients with breast cancer, where patients with high CA 15-3 show good response to aggressive treatments⁸⁹. Serum CA15-3 has been used as a surrogate marker of disease bulk to monitor metastatic breast cancer patients undergoing treatment and for the preclinical detection of tumour recurrence⁸⁹. Elevated levels of CA-15-3 has also been found in patients with other cancers (lung, colorectal, ovarian, pancreatic) and hepatic dysfunction⁴³. The upper limit of normal level of this marker is 25U/ml.

CA19-9 (cancer antigen 19-9) or GICA (gastrointestinal cancer antigen) is a glycolipid with unknown biological function, which was the first successful tumour marker used for serological diagnosis of pancreatic cancer^{90,91}. The nomenclature derives from the monoclonal antibody clone 19-9 that was developed by Koprowski. The concentration of Ca 19-9 in serum has been shown to be a sensitive and specific marker for pancreatic cancer, while its elevated levels in urine have been found in bladder cancer⁹¹. The amount of Ca 19-9 in urine seems to reflect not only the presence of tumour(s) but also the existence of urothelial dysplasia or carcinoma *in situ*⁹². Owing to its high specificity, it plays an important role in the diagnosis, therapeutic monitoring and monitoring of the course of gastrointestinal carcinomas, in particular in the case of pancreatic carcinoma, hepatobiliary carcinoma (carcinoma of the liver, carcinoma of the bile ducts) and carcinoma of the stomach⁹³. However, increased levels of CA 19-9 can also be found in patients with nonmalignant inflammatory diseases, such as cholecystitis and obstructive icterus, cholelithiasis, cholecystolithiasis, acute cholangitis, toxic hepatitis and other liver diseases and therefore should be used with caution^{92,93}. The normal blood levels of CA 19-9 are below 37 U/ml and with this reference level a false positive rate of 20 per cent has been reported in pancreatitis⁹².

Carcinoembryonic antigen (CEA) is a 200 kDa glycoprotein, isolated first by Gold and Freeman in 1965⁹⁴ using an antibody raised in rabbits by injecting an extract of human colonic carcinoma. Elevated levels are found in patients with colorectal, breast, lung, or pancreatic cancer⁹⁵, and also in smokers⁹⁶. The first success in developing a blood test for CEA was in 1965, when the antigen was detected in the blood of some patients with colon cancer. Blood levels of CEA are also elevated in many other cancers such as those of the thyroid, pancreas, liver, stomach, prostate,

ovary, and bladder⁹⁶. Post-operative normalization of serum CEA level has been reported to be a favourable prognostic indicator in lung cancer and the identification of abnormal pre- and post-operative serum CEA levels may be useful in the auxiliary cancer prognosis or post-operative surveillance of colorectal cancer patients⁹⁷.

Human chorionic gonadotrophin (HCG) is a hormone produced normally by the placenta, whose level is elevated in the blood of patients with certain types of testicular and ovarian cancers (germ cell tumours) and choriocarcinoma⁹⁸. In addition, synthesis of free β hCG and its subunits by pelvic carcinomas such as those of the colon, urinary tract, prostate, uterus and vulvo-vagina has also been reported⁹⁸. The presence of increased serum levels of hCG and its metabolites is generally considered to be a sign of a poor prognosis and it has been suggested that β hCG might directly modify the growth of the cancer, leading to a worse outcome. The clinical use of free β hCG as a tumour marker has been limited to a small number of patients owing to a short half life and rapid renal clearance⁹⁹. An elevated blood level of hCG is also be found in the urine of pregnant women and therefore may not be useful as a marker under this condition.

Thyroglobulin (Tg) is a large glycoprotein stored in the follicular colloid of thyroid gland and acts as pro-hormone in the intra-thyroid synthesis of thyroxine (T4) and triiodothyronine (T3). This is another organ-specific tumour marker; associated mainly with patients harbouring differentiated thyroid cancer that arise from the follicle cells (*viz.*, papillary and follicular thyroid cancer) frequently resulting in increased levels of thyroglobulin in the blood¹⁰⁰. The thyroglobulin test is primarily used as a tumour marker to evaluate the effectiveness of treatment for thyroid cancer and to monitor recurrence¹⁰⁰. Tg is generally measured in serum, but measurements can also be made in thyroid cyst fluids and other fluids/tissue obtained by fine needle biopsy of thyroid nodules¹⁰¹. Concomitant measurement of antithyroglobulin is essential as anti-Tg antibodies interfere with the Tg assay and render the assay for thyroglobulin invalid¹⁰². This important factor should be taken into consideration during the evaluation of patient's status. Further, this will not be a very useful marker in tumours which do not release a significant amount of thyroglobulin into the circulation. Although an undetectable level of serum Tg after thyroidectomy and ¹³¹I ablation suggests that patients are free of disease, several studies have shown that a minority of patients, under the influence

of TSH stimulation, have elevated serum Tg¹⁰³ levels and therefore interpretation needs to be made with caution.

Heat shock proteins (HSPs) expression is tailored for particular stress response, with accumulation of denatured proteins as the proximal signal for its induction¹⁰⁴. Heat shock proteins (Hsps) are overexpressed in a wide range of human cancers and are implicated in tumour cell proliferation, differentiation, invasion, metastasis, death, and recognition by the immune system. At present it is unclear as to how the Hsps become overexpressed in cancer; one hypothesis is that the physio-pathological features of the tumour microenvironment (low glucose, pH, and oxygen) stimulate the Hsp induction^{105,106}.

Although Hsp levels are not informative at the diagnostic level, due to overexpression in a wide range of malignant cells and tissues, these are useful biomarkers for carcinogenesis in tissues and are suggestive of the degree of differentiation and the aggressiveness in certain types of cancers¹⁰⁷. Further, the circulating levels of Hsp and anti-Hsp antibodies in cancer patients may be useful in tumour diagnosis. Several Hsp are implicated in the prognosis of specific cancers; most notably Hsp27, whose expression is associated with poor prognosis in gastric, liver, and prostate carcinoma, and osteosarcomas, while Hsp70 is correlated with poor prognosis in breast, endometrial, uterine cervical, and bladder carcinomas. Increased Hsp expression has also been found to predict the response to certain anticancer treatments. While Hsp27 is associated with a poor response to chemotherapy in leukaemia patients, Hsp70 expression predicts a better response to chemotherapy in osteosarcomas¹⁰⁴. Implication of Hsp in tumour progression and response to therapy has led to its successful targeting in therapy by use of Hsps in anticancer vaccines, exploiting their ability to act as immunological adjuvant¹⁰⁷.

Oral fluid contains proteomic signatures that may serve as biomarkers for human diseases such as oral cancer. Therefore, it has been suggested that proteomic analysis of human oral fluid such as whole saliva holds promise as a non-invasive method to identify biomarkers for human oral cancer¹⁰⁸. Most recently detection of five proteins in the saliva of cancer patients has been found to be useful markers of oral cancer with 90 per cent sensitivity and 83 per cent specificity for oral squamous cell carcinoma. These proteins include (*i*) calcium-binding protein MRP14 implicated in several types of

cancer; (ii) CD59 overexpressed on tumour cells that enables them to escape from complement-dependent and antibody-mediated immune responses; (iii) Profilin 1, a protein involved in several signaling pathways with cytoplasmic and nuclear ligands, generally secreted into tumour microenvironments during the early progressive stage of tumorigenesis; and (iv) catalase, a member of the enzymatic antioxidative system, whose level is elevated in many human tumours and involved in carcinogenesis and tumour progression¹⁰⁹. However, long-term studies employing large number of oral cancer patients as well as subjects at high risk of developing oral cancer are needed to validate these potential biomarkers.

Mitochondrial markers Mitochondria typically contain multiple haploid copies of their own genome (16.5 kb), including most components of transcription, translation, and protein assembly. mtDNA is present at 1000-10,000 copies/cell, and the vast majority of these copies are identical (homoplasmic) at birth. Several mutations in the mtDNA, particularly in the D-loop region have been recently found in breast, colon, oesophageal, endometrial, head and neck, liver, kidney, leukemia, lung, melanoma, oral, prostate, and thyroid cancer¹¹⁰. The majority of these somatic mutations are homoplasmic in nature, suggesting that the mutant mtDNA played an active role in tumour formation¹¹¹. By virtue of their clonal nature and high copy numbers in cancer cells, mitochondrial mutations may provide a powerful molecular marker for noninvasive detection of cancer. It may also be useful in early detection, diagnosis, and prognosis of cancer outcome and/or in monitoring response to certain preventive and interventional modalities as well as therapies^{112,113}. Mutated mtDNA has also been detected in the body fluids of cancer patients and indeed is much more abundant than the mutated nuclear p53 DNA¹¹⁴. Since the mitochondrial gene expression signatures of transformed cells have now been identified, development of mitochondrial functional proteomics is expected to identify new markers for early detection and risk assessment, as well as targets for therapeutic intervention¹¹¹. Many advanced techniques are currently available or being newly developed for the studying the mitochondrial proteome *viz.* IP, DiGE, ICAT, SELDI, MALDITOF, Protein/antibody array, *etc.*, are expected to facilitate further this process.

Metabolic biomarker (glucose metabolism)

Enhanced glucose utilization is a prominent and fundamental change in many tumours irrespective of

their histological origin and the nature of mutations, first observed by Warburg^{115,116}. Mechanisms underlying this fundamental alterations in metabolism during carcinogenesis include mutations in the mitochondrial DNA resulting in functional impairment, oncogenic transformation linked upregulation of glycolysis, enhanced expression of metabolic enzymes and adaptation to the hypoxic tumour micro-milieu in case of solid tumours¹¹⁷. Based on these observations, a bioenergetic index of the cell (BEC index) has been suggested that could be used for classification and prognosis of cancers, besides predicting the response to therapy¹¹⁸. Positron emission tomography (PET), which allows non invasive and quantitative analysis of various biologic processes, uses a glucose analogue (2-deoxy-D-glucose) labelled with a positron emitter Fluorine 18; FDG that is partially metabolized and trapped as its phosphate (2-DG-6-P) in the tumour tissue thus localizing the tumour¹¹⁹. The extent of increase in glucose utilization measured by FDG-PET has been correlated with the degree of malignancy in some of the tumours¹²⁰. Glucose utilization is also inversely correlated with treatment response in a number of tumours, while changes in tumour glucose utilization during the first weeks of chemotherapy are significantly correlated with patient outcome^{121,122}. Therefore, glucose utilization appears to be a useful metabolic marker for diagnosis, prognosis and prediction of tumour response to a variety of therapies¹²³.

Therapeutic biomarkers

Cytotoxic chemotherapy and radiotherapy remain the most effective treatments for cancer; however, these can cause serious side effects as these do not often show adequate differential effect between tumour and normal cells. Advances in understanding the molecular basis of cancer made possible by the identification and functional analysis of tumour-specific genetic alterations have opened exciting new opportunities for the design of therapies that specifically target the molecular pathways involved in promoting tumour cell growth and circumvent death pathways like apoptosis.

In the past decade, there have been considerable improvements in the way that human tumours are characterized. Knowledge of cancer at the molecular level has therefore increased greatly, and has catalyzed a shift towards using targeted therapies for cancer. In principle, "targeted therapies" display greater selectivity for tumour cells, and indeed several such therapies have already shown promise in the clinic. These include small molecule drugs that inhibit the activity of protein

tyrosine kinases [*e.g.*, imatinib and erlotinib, targeting ABL and the epidermal growth factor receptor (EGFR), respectively] and neutralizing antibodies that inhibit trans-membrane signaling receptors (*e.g.*, trastuzumab, targeting HER2). Other targeted therapies include drugs that block the activity of molecules in the host microenvironment that support tumour growth (*e.g.*, the antibody bevacizumab, which targets a growth factor that stimulates tumour blood vessel growth). To date, many of these therapies have conferred only modest benefits on patient survival, but refinements in the mode of use of these drugs (*e.g.*, as combination therapies and with biomarker-guided patient selection) are expected to improve their efficacy. Development of clinical tools to identify which patients are most likely to benefit from particular targeted therapies will aid the individualization of molecular targeted therapies thereby enhancing the efficacy of therapy.

Glycolysis

Enhanced glucose dependency is one of the prominent characteristics of most malignant tumours and correlate well with resistance to radio- and chemotherapy^{124,125}. Metabolic status linked alterations in cell signaling related to defense against oxidative stress, redox signaling and damage response pathways, particularly the downregulation of mitochondrial dependent apoptosis in tumour cells with enhanced glucose usage and Hexokinase II levels have been observed¹²⁶. Recent observations in many human tumour cell lines with varying degrees of glycolysis (endogenous and induced) have shown an inverse relationship between the rate of glycolysis (glucose usage and lactate production) and manifestation of damage induced by radiation and chemotherapeutic drugs¹²⁷⁻¹²⁹ similar to the correlations between FDG uptake and responses to therapy clinical responses. These observations have prompted the targeting of this phenotype (elevated glycolysis) as an attractive proposition for developing therapeutics and adjuvant in cancer therapy^{130,131}.

In vitro and *in vivo* studies with several murine and human tumour cells have indeed shown that glycolytic inhibitors like 2-deoxy-D-glucose, 3-bromo-pyruvate *etc.*, are selectively cytotoxic to tumour cells, inducing both growth inhibition and cell death¹³². Unfortunately, therapies using pharmacological inhibitors of glycolysis as a primary therapeutic agent have not produced remarkable results in providing effective local tumour control, besides showing systemic

toxicity, particularly to the central nervous system¹³³. A more promising approach for improving cancer therapy exploiting the inherent differences in glucose metabolism between tumour and normal cells, employs 2-DG as a differential modifier of cellular responses to the widely used therapeutic agents such as radiation and/or cytotoxic drugs¹³⁴⁻¹³⁷. The rationale behind this approach is based on the bioenergetics of cellular damage response pathways including DNA repair, cell proliferation and cell death on the one hand and enhanced glucose dependency in tumour cells on the other. Several studies using *in vitro* and *in vivo* models of tumours have shown that 2-DG selectively sensitizes tumour cells to ionizing radiation, while reducing the damage to normal cells^{138,139}. Since mechanisms underlying cellular responses to damage caused by many anticancer drugs are similar to radiation, it has been suggested that 2-DG has the potential to enhance the efficacy of chemotherapy^{140,141}. Indeed, 2-DG has been found to enhance the damage caused by certain chemotherapeutic drugs *in vitro*¹⁴²⁻¹⁴⁵ and *in vivo*^{146,147}.

Clinical trials in patients with malignant brain tumours (glioblastoma multiforme) using a hypofractionated radiotherapy protocol combined with 2-DG have been very encouraging¹³⁴. Excellent tolerance to the combined treatment, with minimal acute toxicity and late radiation effects as well as increase in survival and significant improvement in the quality of life has been reported^{134, 135}.

Mammalian target of rapamycin (mTOR)

This is an evolutionarily conserved serine-threonine protein kinase that belongs to the PI3K [phosphoinositide 3-kinase (PI3K)-related kinase] family, and plays an important role in regulating cell growth and proliferation¹⁴⁸. Upon activation, mTOR increases the phosphorylation levels of its downstream targets that include p70S6K and 4EBP1, which leads to increased levels of translation, ribosome biogenesis, and reorganization of the actin cytoskeleton and inhibition of autophagy. As a result, mTOR activation promotes cell growth and proliferation, whereas mTOR inhibition stops cell growth and initiates catabolic processes, including autophagy¹⁴⁹.

The phosphatidylinositol-3-OH kinase (PI(3)K)–PTEN–mTOR signaling pathway is aberrantly activated in many tumours, leading to dysregulation of cell growth and proliferation¹⁴⁸. Activation of this pathway can be assessed by biomarkers such as loss of PTEN mRNA or protein production in tumour tissue.

Biochemical inhibition of mTOR by rapamycin can be assessed by biomarkers such as the abundance of the phosphorylated form of the ribosomal protein S6, and its therapeutic effects on tumour cells can be assessed by the proliferation marker Ki-67¹⁵⁰.

A number of mTOR inhibitors have potent antiproliferative properties which make them useful for cancer chemotherapy, particularly of advanced solid tumours¹⁵¹. It has surprisingly been found that S6 40S ribosomal protein (otherwise known as S6) is a useful biomarker which is predictive of sensitivity of proliferative diseases to treatment with an mTOR inhibitor. In particular, it has been found that the phosphorylation state of S6 correlates well with sensitivity to mTOR inhibitors. mTOR inhibitors are more likely to show a significant antiproliferative effect when used to treat cancer cell lines showing higher levels of expression of phosphorylated S6. Moreover, the method may be used to select an appropriate dose of an mTOR inhibitor in order to individualize therapy for each patient¹⁵².

Telomerase

Telomeres are tracts of repetitive DNA (TTAGGG/AATCCC for human telomeres) that protect chromosomes from degradation and loss of essential genes. Under normal circumstances, telomeres progressively shorten in most human cells with each cycle of cell division and the length in adult human tissues is approximately half that of the new born. Telomerase belongs to a class of enzymes known as reverse transcriptases that use RNA as a template for creating DNA and it contains both RNA and protein components. The enzyme ensures the maintenance of telomere and thereby protecting the cell from degradation and death¹⁵³. Since telomerase is found in nearly 90 per cent of human cancers and is responsible for indefinite growth of cancer cells^{154,155}, it has been a target for anticancer therapeutics that turn-off telomerase and thereby inhibit tumour growth. The levels of telomerase are also elevated in stem cells allowing unlimited division necessary for the repair of damaged and worn out tissues. Most human tumours not only express telomerase but interestingly also have very short telomeres. Telomerase is one of the best markers for human cancer, associated with only malignant tumours and not the benign lesions making it a diagnostic marker as well as an ideal target for chemotherapy¹⁵⁶⁻¹⁵⁸.

In normal cells, telomerase is sequestered in an area of the cell nucleus called the nucleolus, away from

the chromosomes. The enzyme is released only when needed during cell division, and then returns quickly to the nucleolus thereafter. In cancer cells, however, telomerase is found throughout the cell, implying that the telomerase-shuttling system is impaired. Identification and manipulation of proteins normally involved in telomerase transfer could prove to be useful targets for anti-telomerase therapies¹⁵⁹. Currently two clinical trials; one using a vaccine (GRNVAC1) and the other a lipidated drug (GRN163L) are underway to evaluate the efficacy of telomerase inhibitors¹⁶⁰.

p53

The *p53* gene is one of the tumour suppressor genes that normally prevent uncontrolled multiplication of abnormal cells and experimental findings from the last two decades have established a crucial role for wild-type *p53* in intrinsic tumour suppression^{161,162}. Upon stimulation (*e.g.*, by moderate levels of DNA damage), *p53* activates molecular processes that delays the cell cycle progression of proliferating cells and simultaneously stimulating DNA repair processes^{163,164}. On the other hand, higher level of damage has been found to activate *p53* mediated cell death pathway (typically apoptosis), a mechanism that is purported to be responsible for the prevention of carcinogenesis. During malignant transformation *p53* or *p53*-pathway related molecules are disabled most often and a mutant form of *p53* may not only negate the wild type *p53* function but plays additional role in tumour progression¹⁶⁵. Nearly 50 per cent of all human tumours carry a mutated *p53* gene¹⁶⁵. Clinical studies in patients with various types of cancer have shown that certain mutations in the *p53* gene are significant predictor of resistance to therapy¹⁶³.

Although *p53* is not a typical cancer-specific antigen, its central role in the control of cell growth and apoptosis and frequent mutations in tumours make *p53* a unique target for cancer therapy. Radiation and many of the anticancer drugs damage the DNA of cancer cells, triggering the action of the *p53* leading to apoptosis. Hence, an intact wild type *p53* gene is essentially required to stimulate programmed cell death of a cancer cell in response to treatment. Investigations in several types of cancer have shown that the *p53* gene is a potentially useful biomarker for predicting prognosis and patient's response to therapy¹⁶⁵. Experimental evidences show that either mutation in the gene or overexpression of the *p53* protein can be used to predict many aspects of

prognosis and outcome of patients with various type of cancer.

Among the different approaches targeting p53, replacement gene therapies that have been explored extensively in recent years aims at restoration of p53 function in cancer cells by introduction of exogenous p53. Various protocols and vectors have been employed, including retroviruses, adenoviruses and vaccinia-derived vectors¹⁶⁶. Recent studies have focused on adenoviral vectors, with Ad-p53, adenovirus serotype 5 carrying *wt p53* genes, as a model example. Although preliminary results were promising, recent clinical data failed to demonstrate anti-tumour activity in patients and some trials have indeed been discontinued¹⁶⁷. New trials aim at a combination of gene transfer with chemotherapy or radiotherapy. In addition to the strategy of p53 reactivation in tumours, modulation of p53 activity in normal cells may protect them from the side effects of chemotherapy or radiotherapy. Several new compounds targeting p53 have entered clinical trials and therefore, p53-oriented therapy will be one of the major areas of intense investigations in the coming years. However, the approach of restoring of p53 function in tumour cells has been nearly questioned after a few contradictory results, which shows that prostate cancer cells are protected from ionizing radiation-induced DNA damage through activation of p53¹⁶⁸ and cells transformed with oncogenic tyrosine kinase BCR/ABL may actually benefit from activation of p53 upon DNA damage¹⁶⁵. These observations have led to the reexamination of restoring p53 function in tumors as a therapeutic strategy.

Tyrosine kinase

Tyrosine kinases are a class of enzymes that regulate multiple cellular processes by acting primarily as important transducers of extracellular signals influencing diverse functions such as cell growth, differentiation, migration, and apoptosis that contribute to tumour development and progression. Many human tumours display aberrant activation of tyrosine kinases caused by genetic alterations that could be related to the malignant transformation¹⁶⁹. The erbB or HER family of transmembrane tyrosine kinase receptors, especially receptors erbB1 (or EGFR) and erbB2 (or Her2/neu), has been identified as an important therapeutic target in a number of cancers. Her2/neu, is overexpressed in nearly 30 per cent of patients with aggressive breast cancer, while EGFR is overexpressed in several solid tumours¹⁷⁰. Therefore, targeting protein tyrosine kinases

as a therapeutic strategy has been very attractive and results from the recent clinical studies are indeed quite encouraging. Current approaches include blocking kinase-substrate interaction, inhibiting the enzyme's adenosine triphosphate (ATP) binding site and blocking extracellular tyrosine kinase receptors on tumour cells. Several tyrosine kinase inhibitors (TKIs) (*viz.*, gefitinib and trastuzumab) have already been approved as anti-cancer agents.

Histone deacetylases (HDACs)

Acetylation of proteins orchestra the dynamic interplay between various processes like repair of DNA damage, cell cycle arrest and apoptosis determining the cellular response to radiation and various chemotherapeutic drugs. This acetylation is catalyzed by histone acetylases (HATs) that uses acetyl-CoA as substrate and the acetyl group is transferred to the ϵ amino group of certain lysine side chains within histones N-terminal tails and other nuclear receptor proteins thereby regulating chromatin remodeling and gene expression¹⁷¹. Chromatin remodeling during the regulation of gene expression is orchestrated by a concerted action of HATs and HDACs that condenses and decondenses the chromatin structure by acetylating and deacetylating histones and other nuclear receptor proteins. Further, HDACs appear to be closely associated with oncogenesis by regulating the expression of certain tumour suppressor genes leading to excessive proliferation and tumorigenesis¹⁷². HDAC have recently been among some of the attractive targets for cancer therapeutics, and HDAC inhibitors with diversified structures have indeed shown promising anti-tumour activity (cell cycle arrest, cellular differentiation and apoptosis) both *in vitro*¹⁷³ and *in vivo*¹⁷⁴. Many of the HDAC inhibitors are currently under clinical investigation in a number of haematological malignancies and solid tumours^{175,176}. Further, HDAC inhibitors are also being investigated as adjuvant together with other anti-cancer therapeutics¹⁷⁷. It appears therefore, that HDAC inhibitors with pleiotropic actions in modulating multiple genes, signaling pathways and biological features of malignancy are useful in the treatment of cancers with multiple oncogenic abnormalities targeting the protein acetylation involved in the regulation of cell signaling¹⁷⁸.

PINI

It is well known that the functional status of many proteins is regulated by kinase mediated phosphorylation

and other post-translational modifications. Recently, regulation of proteins beyond phosphorylation has been unraveled, which is in the form *Cis* and *Trans* isomerization (a post-phosphorylation event) of phosphoserine/threonine - proline peptide bonds at selective sites catalyzed by peptidyl-prolyl isomerase (PPIase), Pin1^{179,180}. These conformational changes can have profound effect on the function of proteins, modulating their activity, phosphorylation status, protein-protein interactions, subcellular localization and stability. Overexpression of Pin1 has been reported in human breast cancer cell lines and tissues, and its expression closely correlates with the level of cyclin D1 (important cyclin required for cell proliferation) in tumours¹⁸¹. Pin1 overexpression not only confers transforming properties on normal mammary epithelial cells, but also enhances transformed phenotypes of Neu/Ras-transformed mammary epithelial cells and implicated in mitotic regulation¹⁸². In contrast, inhibition of Pin1 suppresses the Neu- and Ras induced transformed phenotypes or induces tumour cells into mitotic arrest and apoptosis^{182,183}. Pin1 opens a new target for the development of specific therapeutics and has received greater attention as phosphorylated p53 is among the known substrates of Pin1^{184,185}. Inhibition of Pin 1 through various approaches, such as mutations, deletions or expression of antisense, induces mitotic arrest and apoptosis in tumour cell lines¹⁸⁶. It appears that Pin1 can be used as a diagnostic marker for the detection of the cancer or to stage the disease, albeit in only certain types of cancers.

Recent studies have shown that treatment of cells with pin1 inhibitor juglone delays the growth of various tumour cell lines¹⁸⁷, suggesting that inhibition of pin1 can be used as an approach for inhibiting tumour growth. We have recently identified potential Pin1 sites on topoisomerase II α (a vital nuclear enzyme and mitotic protein) and shown that the two proteins functionally interact with each other resulting in the activation of topo II α ^{188,189}. Moreover, using inhibitors of topo II α (etoposide) and Pin1 (Juglone), we have shown that the combination (etoposide and juglone) may improve the therapeutic potential^{188,190}. Pin1 appears to be an attractive target for diagnosis and therapy. Further understanding on the role of Pin1 in tumourigenesis is required before its use as a target for developing antagonists ensuring specificity, selectivity and safety.

Conclusion

Discovery and clinical application of new biomarkers, is expected to play a significant role in reshaping life

science research and life science industry, thereby profoundly influencing the detection and treatment of many diseases and cancer in particular. Clinical oncology is poised to enter a new era in which cancer detection, diagnosis, and treatment will be guided increasingly by the molecular attributes of the individual patient, acquired from several different sources *viz.*, tumour tissue, host cells/tissues that influence tumour behaviour and body fluids. The resultant panel of biomarkers will not only help the detection and diagnosis, but also answer fundamental questions about biologic behaviour of tumours, resistance to therapy and sensitivity to therapy facilitating individualization of therapy, besides identifying individuals predisposed to cancer. The future of cancer therapy lie in the use of biomarkers that offer the potential to identify and treat cancer years before it is either visible or symptomatic. Exploring the presence of such markers that does not require the tumour tissue to detect them, but are secreted by cancer cells into the blood stream will not only facilitate easy detection without even minimal surgical procedure, but will also be candidates for population based screening.

Contemporary as well as upcoming genomic and proteomic technologies are quite promising in identifying new biomarkers, which can significantly enhance the efficacy of cancer management by facilitating the individualization of therapy targeting the patient specific molecular lesions and also by providing tools for predicting/monitoring of therapeutic response. Although the current understanding of signaling pathways has identified specific targets for developing newer drugs and therapeutic strategies, a comprehensive understanding of how the complex signaling networks function in intact cell is still required, to evolve strategies based on the genetic alterations in individual cancers.

Future challenges in the biomarkers using genomic and proteomic diagnostic technology include the development of complex mathematical algorithms to handle simultaneous analysis of many parameters (perhaps up to thousand even) to aid the diagnosis instead of a single parameter. Further, issues regarding quality control methods and procedures also need to be developed for using these markers with reliability and reproducibility. A comprehensive understanding of the relevance of each biomarker will be very important to efficiently diagnose the disease and provide appropriate direction in the multiple therapeutic alternatives currently available that is likely to benefit the unfortunate patients.

References

- Srinivas PR, Kramer BS, Srivastava S. Trends in biomarker research for cancer detection. *Lancet Oncol* 2001; 2 : 698-704.
- Cho WSC. Contribution of oncoproteomics to cancer biomarker discovery. *Mol Cancer* 2007; 6 : 25.
- Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell* 2000; 100 : 57-70.
- Bayli SB, Ohm JE. Epigenetic gene silencing in cancer - a mechanism for early oncogenic pathway addiction? *Nature Rev Cancer* 2006; 6 : 107-17.
- Jaenisch R, Bird A. Epigenetic regulation of gene expression: how the genome integrates intrinsic and environmental signals. *Nature Genet* 2003; 33 : 245-54.
- Ludwig JA, John N. Weinstein biomarkers in cancer staging, prognosis and treatment selection. *Nat Rev Cancer* 2005; 5 : 845-56.
- Weissleder AR, Ntziachristos V. Shedding light onto live molecular targets. *Nat Med* 2003; 9 : 123-8.
- Sidransky BD. Emerging molecular markers of cancer. *Nat Rev Cancer* 2002; 2 : 210-9.
- Vogelstein CB, Kinzler KW. Cancer genes and the pathways they control. *Nat Med* 2004; 10 : 789-99.
- Sawyers CL. The cancer biomarker problem. *Nature* 2008; 452 : 548-52.
- Dwarakanath BS, Manogaran PS, Das S, Das BS, Jain V. Heterogeneity in DNA content and proliferative status of human brain tumors. *Indian J Med Res* 1994; 100 : 279-86.
- Whitfield ML, George LK, Grant GD, Perou CM. Common markers of proliferation. *Nat Rev Cancer* 2006; 6 : 99-106.
- Zarbo RJ, Nakhleh RE, Brown RD, Kubus JJ, Ma CK. Prognostic significance of DNA ploidy and proliferation in 309 colorectal carcinomas as determined by two-color multiparametric DNA flow cytometry. *Cancer* 2000; 79 : 2073-86.
- Bishop JM. The molecular genetics of cancer. *Science (Wash. DC)* 1987; 235 : 305-11.
- Dunning AM, Healey CS, Pharoah PDP, Teare DM, Ponder BAJ, Easton DF. A systematic review of genetic polymorphisms and breast cancer risk. *Cancer Epidem Biomarkers Prevent* 1999; 8 : 843-54.
- Toru H, Masaharu Y, Shinji T, Kazuaki C. Genetic polymorphisms and head and neck cancer risk. *Int J Oncol* 2008; 32 : 945-73.
- Ignatiadis M, Xenidis N, Perraki M, Apostolaki S, Politaki E, Kafousi M, et al. Different prognostic value of Cytokeratin-19 mRNA positive circulating tumor cells according to estrogen receptor and HER2 status in early-stage breast cancer. *J Clin Oncol* 2007; 25 : 5194-202.
- Delys L, Detours V, Franc B, Thomas G, Bogdanova T, Tronko M, et al. Gene expression and the biological phenotype of papillary thyroid carcinomas. *Oncogene* 2007; 26 : 7894-903.
- Jeffery PL, Herington AC, Chopin LK. Ghrelin and a novel ghrelin isoform have potential autocrine/paracrine roles in hormone-dependent cancer. *Endocrine Abstracts* 2003; 6 : 38.
- Alison MR, Hunt T, Forbes SJ. Minichromosome maintenance (MCM) proteins may be pre-cancer markers. *Gut* 2002; 50 : 290-1.
- Forsburg SL. Eukaryotic MCM proteins: beyond replication initiation. *Microbiol Mol Biol Rev* 2004; 68 : 109-31.
- Carter SL, Negrini M, Baffa R, Gillium DR, Rosenberg AL, Schwartz GF, et al. Loss of heterozygosity at 11q22-q23 in breast cancer. *Cancer Res* 1994; 54 : 6270-4.
- Thor AD, Moore DH, Edgerton SM, Kawasaki ES, Reihnsaus E, Lynch HT, et al. Accumulation of p53 tumor suppressor gene protein: an independent marker of prognosis in breast cancers. *J Natl Cancer Inst* 1992; 84 : 845-55.
- Ngan BY, Chen L, Weiss LM, Warnke RA, Cleary ML. Expression in non-Hodgkin's lymphoma of the bcl-2 protein associated with the t(14;18) chromosomal translocation. *N Engl J Med* 1988; 318 : 1638-44.
- Arzimanoglou I, Gilbert F, Barger HR. Microsatellite instability in human solid tumors. *Cancer* 1998; 82 : 1808-20.
- Weber JL, May PE. Abundant class of human DNA polymorphisms which can be typed using polymerase chain reaction. *Am J Hum Genet* 1989; 44 : 388-96.
- Arnold K. Biomarker for esophageal cancer found in bloodstream. *J Nat Cancer Inst* 2000; 92 : 1787.
- Fearnhead NS, Britton MP, Bodmer WF. The ABC of APC. *Hum Mol Genet* 2001; 10 : 721-33.
- Puig PL, Bérout C, Soussi T. APC gene: database of germline and somatic mutations in human tumors and cell lines. *Nucleic Acids Res* 1998; 26 : 269-70.
- Egger G, Liang G, Aparicio A, Jones PA. Epigenetics in human disease and prospects for epigenetic therapy. *Nature* 2004; 429 : 457-63.
- Jones PA, Takai D. The role of DNA methylation in mammalian epigenetics. *Science* 2001; 293 : 1068-70.
- Herman JG, Baylin SB. Gene silencing in cancer in association with promoter hypermethylation. *N Engl J Med* 2003; 349 : 2042-54.
- Ehrlich M. DNA methylation in cancer: too much, but also too little. *Oncogene* 2002; 21 : 5400-13.
- Kim H, Kwon YM, Kim JS, Lee H, Park JH, Shim YM, et al. Tumor-specific methylation in bronchial lavage for the early detection of non-small-cell lung cancer. *J Clin Oncol* 2004; 22 : 2363-70.
- Battagi C, Uzzo RG, Dulaimi E, Ibanez de Caceres I, Krassenstein R, Al-Saleem T, et al. Promoter hypermethylation of tumor suppressor genes in urine from kidney cancer patients. *Cancer Res* 2003; 63 : 8695-9.
- Nakayama H, Hibi K, Takase T, Yamazaki T, Kasai Y, Ito K, et al. Molecular detection of p16 promoter methylation in the serum of recurrent colorectal cancer patients. *Int J Cancer* 2003; 105 : 491-3.
- Belinsky SA. Gene-promoter hypermethylation as a biomarker in lung cancer. *Nat Rev Cancer* 2004; 4 : 707-17.
- Esteller M, Garcia-Foncillas J, Andion E, Goodman SN, Hidalgo OF, Vanaclocha V, et al. Inactivation of the DNA-repair gene MGMT and the clinical response of gliomas to alkylating agents. *N Engl J Med* 2000; 343 : 1350-4.

39. Ring A, Smith IE, Dowsett M. Circulating tumour cells in breast cancer. *Lancet Oncol* 2004; 5 : 79-88.
40. Cristofanilli M, Budd GT, Ellis MJ, Stopeck A, Matera J, Miller MC, *et al.* Circulating tumor cells predict progression free survival and overall survival in metastatic breast cancer. *N Engl J Med* 2004; 351 : 781-91.
41. Shaffer DR, Leversha MA, Danila DC, Lin O, Gonzalez-Espinoza R, Gu B, *et al.* Circulating tumor cell analysis in patients with progressive castration resistant prostate cancer. *Clin Cancer Res* 2007; 13 : 2023-9.
42. Cristofanilli M, Budd GT, Ellis MJ, Stopeck A, Matera J, Miller MC, *et al.* Presence of circulating tumor cells (CTC) in metastatic breast cancer (MBC) predicts rapid progression and poor prognosis. *J Clin Oncol* 2005; 23 : 524.
43. Sakaguchi S. Naturally arising CD4⁺ regulatory T cells for immunologic self-tolerance and negative control of immune responses. *Ann Rev Immunol* 2004; 22 : 531-62.
44. Maloy KJ, Powrie F. Regulatory T cells in the control of immune pathology. *Nat Immunol* 2001; 2 : 816-22.
45. Simon AK, Jones E, Richards H, Wright K, Betts G, Godkin A, *et al.* Regulatory T cells inhibit Fas ligand-induced innate and adaptive tumour immunity. *Eur J Immunol* 2007; 37 : 758-67.
46. Liyanage UK, Moore TT, Joo H-G, Tanaka Y, Herrmann V, Doherty G, *et al.* Prevalence of regulatory T cells is increased in peripheral blood and tumor microenvironment of patients with pancreas or breast adenocarcinoma. *J Immunol* 2002; 169 : 2756-61.
47. Siddiqui SA, Frigola X, Bonne-Annee S, Mercader M, Kuntz SM, Krambeck AE, *et al.* Tumor-infiltrating Foxp3⁺ CD4⁺CD25⁺ T cells predict poor survival in renal cell carcinoma. *Clin Cancer Res* 2007; 13 : 2075-81.
48. Woo EY, Yeh H, Chu CS, Schlienger K, Carroll RG, Riley JL, *et al.* Regulatory T cells from lung cancer patients directly inhibit autologous T cell proliferation. *J Immunol* 2002; 168 : 4272-6.
49. Wolf AM, Wolf D, Steurer M, Gastl G, Gonsilius E, Grubeck-Loebenstien B. Increase of regulatory T cells in the peripheral blood of cancer patients. *Clin Cancer Res* 2003; 9 : 606-12.
50. Viguier M, Lemaître F, Verola O, Cho MS, Gorochoy G, Dubertret L, *et al.* Ferradini Foxp3 expressing CD4⁺CD25⁺ high regulatory T cells are overrepresented in human metastatic melanoma lymph nodes and inhibit the function of infiltrating T cells. *J Immunol* 2004; 173 : 1444-53.
51. Ormandy LA, Hillemann T, Wedemeyer H, Manns MP, Greten TF, Korangy F. Increased populations of regulatory T cells in peripheral blood of patients with hepatocellular carcinoma. *Cancer Res* 2005; 65 : 2457-64.
52. Joosten SA, Ottenhoff THM. Human CD4 and CD8 regulatory T cells in infectious diseases and vaccination. *Hum Immunol* 2008; 69 : 760-70.
53. Curiel T, Coukos G, Zou L, Alvarez X, Cheng P, Mottram P, *et al.* Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nature Med* 2004; 10 : 942-9.
54. Farooque A, Singh S, Adhikari JS, Dwarakanath BS. Role of T regulatory cells (CD4⁺CD25⁺FoxP3⁺) and Th1, Th2 and Th3 cytokines in the radiosensitization of Ehrlich ascites tumor by the glycolytic inhibitor 2-deoxy-D- glucose (2-DG). *XXIV International Congress on Cytometry in the Age of Systems Biology*, May 17-21, 2008: Budapest, Hungary; p.156.
55. Schreiber TH. The use of FoxP3 as a biomarker and prognostic factor for malignant human tumors. *Cancer Epidemiol Biomarkers Prevent* 2007; 16 : 1931-4.
56. Clarke MF, Dick JE, Dirks PB, Eaves CJ, Jamieson CH, Jones DL, *et al.* Cancer stem cells - perspectives on current status and future directions: AACR workshop on cancer stem cells. *Cancer Res* 2006; 66 : 9339-44.
57. Clarke MF, Fuller M. Stem cells and cancer: two faces of eve. *Cell* 2006; 124 : 1111-5.
58. Takaishi S, Okumura T, Wang TC. Gastric cancer stem cells. *J Clin Oncol* 2008; 26 : 2876-82.
59. Bao S, Wu Q, McLendon RE, Hao Y, Shi Q, Hjelmeland AB, *et al.* Glioma stem cells promote radioresistance by preferential activation of the DNA damage response. *Nature* 2006; 444 : 756-60.
60. Singh SK, Hawkins C, Clarke ID, Squire JA, Bayani J, Hide T, *et al.* Identification of human brain tumour initiating cells. *Nature* 2004; 432 : 396-401.
61. Chen CJ, Wang LY, Yu MW. Epidemiology of hepatitis B virus infection in the Asia-Pacific region. *J Gastroenterol Hepatol* 2000; 15 (Suppl): E3-E6.
62. Kirk GD, Bah E, Montesano R. Molecular epidemiology of human liver cancer: Insights into etiology, pathogenesis and prevention from The Gambia, West Africa. *Carcinogenesis* 2006; 27 : 2070-82.
63. Feitelson MA The pathogenesis of chronic hepatitis B virus infection. *Bull Inst Pasteur* 1998; 96 : 227-36.
64. Garland SM. Can cervical cancer be eradicated by prophylactic HPV vaccination? Challenges to vaccine implementation. *Indian J Med Res* 2009; 130 : 311-21.
65. Bharadwaj M, Hussain S, Nasare V, Das BC. HPV & HPV vaccination: Issues in developing countries. *Indian J Med Res* 2009; 130 : 327-33.
66. Shukla S, Bharti AC, Mahata S, Hussain S, Kumar R, Hedau S, *et al.* Infection of human papillomaviruses in cancers of different human organ sites *Indian J Med Res* 2009; 130 : 222-33.
67. Kreimer AR, Clifford GM, Snijders PJ, Castellsagué X, Meijer CJ, Pawlita M, *et al.* HPV16 semiquantitative viral load and serologic biomarkers in oral and oropharyngeal squamous cell carcinomas. *Int J Cancer* 2005; 115 : 329-32.
68. Thompson MP, Kurzrock R. Epstein-Barr virus and cancer. *Clin Cancer Res* 2004; 10 : 803-21.
69. Lin JC, Wang WY, Chen KY, Wei YH, Liang WM, Jan JS, *et al.* Quantification of plasma Epstein-Barr virus DNA in patients with advanced nasopharyngeal carcinoma. *N Engl J Med* 2004; 350 : 2461-70.
70. Gandhi MK, Lambley E, Burrows J, Dua U, Elliott S, Shaw PJ, *et al.* Plasma Epstein-Barr Virus (EBV) DNA is a biomarker for EBV-positive Hodgkin's lymphoma. *Clin Cancer Res* 2006; 12 : 460-4.
71. Stephan C, Jung K, Diamandis EP, Rittenhouse HG, Lein M, Loening SA. Prostate-specific antigen, its molecular forms, and other kallikrin markers for detection of prostate cancer. *Urology* 2002; 59 : 2-8.

72. Papsidero LD, Wang MC, Valenzuela LA, Murphy GP, Chu TMA. Prostate antigen in sera of prostatic cancer patients. *Cancer Res* 1980; 40 : 2428-32.
73. Thompson I, Pauler D, Goodman P, Tangen C, Lucia M, Parnes H, *et al.* Prevalence of prostate cancer among men with a prostate-specific antigen level < or =4.0 ng per milliliter. *N Engl J Med* 2004; 350 : 2239-46.
74. Denmeade SR, Isaacs JT. The role of prostate-specific antigen in the clinical evaluation of prostatic disease. *BJU Int* 2004; 93 : 10-5.
75. Yousef GM, Diamandis EP. An overview of the kallikrein gene families in humans and other species: Emerging candidate tumour markers. *Clin Biochem* 2003; 36 : 443-52.
76. Brawer MK. Radiation therapy failure in prostate cancer patients: risk factors and methods of detection. *Rev Urol* 2002; 4 : 2-S1.
77. Abelev GI. Alpha-fetoprotein in ontogenesis and its association with malignant tumors. *Adv Cancer Res* 1971; 14 : 295-358.
78. Paul SB, Gulati MS, Sreenivas V, Madan K, Gupta AK, Mukhopadhyay S, *et al.* Evaluating patients with cirrhosis for hepatocellular carcinoma: value of clinical symptomatology, imaging and alpha-fetoprotein. *Oncology* 2007; 72 : 117-23.
79. Ball D, Rose E, Alpert E. Alpha-fetoprotein levels in normal adults. *Am J Med Sci* 1992; 303 : 157-9.
80. O'Brien TJ, Tanimoto H, Konishi I, Gee M. More than 15 years of CA 125: what is known about the antigen, its structure and its function. *Int J Biol Markers* 1998; 13 : 188-95.
81. Bast RC Jr, Urban N, Shridhar V, Smith D, Zhang Z, Skates S, *et al.* Early detection of ovarian cancer: promise and reality. *Cancer Treat Res* 2002; 107 : 61-97.
82. Meden H, Fattahi-Meibodi A. CA 125 in benign gynecological conditions. *Int J Biol Markers* 1998; 13 : 231-7.
83. Bast RC, Xu FJ, Yu YH, Barnhill S, Zhang Z, Mills GB. CA 125: the past and the future. *Int J Biol Markers* 1998; 13 : 179-87.
84. Grover S, Koh H, Weideman P, Quinn MA. The effect of the menstrual cycle on serum CA 125 levels: a population study. *Am J Obstet Gynecol* 1992; 167 : 1379-81.
85. Van der Burg ME, Lammes FB, van Putten WL, Stoter G. Ovarian cancer: the prognostic value of the serum half-life of CA125 during induction chemotherapy. *Gynecol Oncol* 1988; 30 : 307-12.
86. Rustin GJS, Bast RC Jr., Kelloff GJ, Barrett JC, Carter SK, Nisen PD, *et al.* Use of CA-125 in clinical trial evaluation of new therapeutic drugs for ovarian cancer. *Clin Cancer Res* 2004; 10 : 3919-26.
87. Duffy MJ. CA 15-3 and related mucins as circulating markers in breast cancer. *Ann Clin Biochem* 1999; 36 : 579-86.
88. Park BW, Oh JW, Kim JH, Park SH, Kim KS, Kim JH, *et al.* Preoperative CA 15-3 and CEA serum levels as predictor for breast cancer outcomes. *Annals Oncol* 2008; 19 : 675-81.
89. Shering S, Sherry F, McDermott E, Higgins NO, Duffy MJ. Preoperative CA 15-3 concentrations predict outcome in breast cancer. *Cancer* 1998; 83 : 2521-7.
90. Koprowski H, Steplewski Z, Mitchell K, Herlyn M, Herlyn D, Fuhrer P. Colorectal carcinoma antigens detected by hybridoma antibodies. *Somatic Cell Genet* 1979; 5 : 957-71.
91. Casetta G, Piana P, Cavallini A, Vottero M, Tizzani A. Urinary levels of tumor associated antigens (CA 19-9, TPA and CEA) in patients with neoplastic and non-neoplastic urothelial abnormalities. *Br J Urol* 1993; 72 : 60-4.
92. Duffy MJ. CA 19-9 as a marker for gastrointestinal cancers: a review. *Ann Clin Biochem* 1998; 35 : 364-70.
93. Uygur-Bayramicli O, Dabak R, Orbay E, Dolapcioglu C, Sargin M, Kilicoglu G, *et al.* Type 2 diabetes mellitus and CA 19-9 levels. *World J Gastroenterol* 2007; 13 : 5357-9.
94. Gold P, Freeman SO. Demonstration of tumor-specific antigens in human colonic carcinoma by immunological tolerance and absorption techniques techniques. *J Exp Med* 1965; 121 : 439-62.
95. Alaoui-Jamali MA, Xu Y. Proteomic technology for biomarker profiling in cancer: an update. *J Zhejiang Univ Sci B* 2006; 7 : 411-20.
96. Khan MS, Chaouachi K, Mahmood R. Hookah smoking and cancer: carcinoembryonic antigen (CEA) levels in exclusive/ever hookah smokers. *Harm Reduction J* 2008; 5 : 19.
97. Wang JY, Lu CY, Chu KS, Ma CJ, Wu DC, Tsai HL, *et al.* Prognostic significance of pre- and postoperative serum carcinoembryonic antigen levels in patients with colorectal cancer. *Eur Surg Res* 2007; 39 : 245-50.
98. Cole LA. Immunoassay of human chorionic gonadotropin, its free subunits, and metabolites. *Clin Chem* 1997; 43 : 2233-43.
99. Kurtzman J, Wilson H, Rao CV. A proposed role for hCG in clinical obstetrics. *Sem Reprod Med* 2001; 19 : 63-8.
100. Mazzaferri EL, Robbins RJ, Spencer CA, Braverman LE, Pacini F, Wartofsky L, *et al.* A consensus report of the role of serum thyroglobulin as a monitoring method for low-risk patients with papillary thyroid carcinoma. *J Clin Endocrinol Metab* 2003; 88 : 1433-41.
101. Pacini F, Fugazzola L, Lippi F, Ceccarelli C, Centoni R, Miccoli P, *et al.* Detection of thyroglobulin in fine needle aspirates of non thyroidal neck masses: a clue to the diagnosis of metastatic differentiated thyroid cancer. *J Clin Endocrinol Metab* 1992; 74 : 1401-4.
102. Baloch Z, Carayon P, Conte-Devolx B, Demers LM, Feldt-Rasmussen U, Henry JF, *et al.* Laboratory medicine practice guidelines. Laboratory support for the diagnosis and monitoring of thyroid disease. *Thyroid* 2003; 13 : 3-126.
103. Kloos RT, Mazzaferri EL. A single recombinant human thyrotropin-stimulated serum thyroglobulin measurement predicts differentiated thyroid carcinoma metastases three to five years later. *J Clin Endocrinol Metab* 2005; 90 : 5047-57.
104. Voellmy R. On mechanisms that control heat shock transcription factor activity in metazoan cells. *Cell Stress Chaperones* 2004; 9 : 122-33.
105. Pinashi-Kimhi O, Michalowicz D, Ben-Zeev A, Oren M. Specific interaction between the p53 cellular tumour antigen and major heat shock proteins. *Nature* 1986; 320 : 182-5.
106. Daniels GA, Sanchez-Perez L, Diaz RM, Kottke T, Thompson J, Lia M, *et al.* A simple method to cure established tumors by inflammatory killing of normal cells. *Nat Biotechnol* 2004; 22 : 1125-32.

107. Cioccal DR, Stuart K. Calderwood Heat shock proteins in cancer: diagnostic, prognostic, predictive, and treatment implications. *Cell Stress Chaperones* 2005; 10 : 86-103.
108. Hu S, Yu T, Xie Y, Yang Y, Li Y, Zhou X, *et al*. Discovery of oral fluid biomarkers for human oral cancer by mass spectrometry. *Cancer Genomics Proteomics* 2007; 4 : 55-64.
109. Hu S, Arellano M, Boonthung P, Wang J, Zhou H, Jiang J, *et al*. Salivary proteomics for oral cancer biomarker discovery. *Clin Cancer Res* 2008; 14 : 6246-52.
110. Jakupciak JP, Wang W, Markowitz ME, Ally D, Coble M, Srivastava S, *et al*. Mitochondrial DNA as a cancer biomarker. *J Mol Diagn* 2005; 7 : 258-67.
111. Maitra A, Cohen Y, Gillespie SE, Mambo E, Fukushima N, Hoque MO, *et al*. The Human MitoChip: a highthroughput sequencing microarray for mitochondrial mutation detection. *Genome Res* 2004; 14 : 812-9.
112. Kim MM, Clinger JD, Masayeva BG, Ha PK, Zahurak ML, Westra WH, *et al*. Mitochondrial DNA quantity increases with histopathologic grade in premalignant and malignant head and neck lesions. *Cancer Res* 2004; 10 : 8512-5.
113. Lièvre A, Blons H, Houllier AM, Laccourreye O, Brasnu D, Beaune P, *et al*. Clinicopathological significance of mitochondrial D-Loop mutations in head and neck carcinoma. *Br J Cancer* 2006; 94 : 692-7.
114. Fliiss MS, Usadel H, Caballero OL, Li WU, Buta MR, Eleff SM, *et al*. Facile detection of mitochondrial DNA mutations in tumors and bodily fluids. *Science* 2000; 287 : 2017-9.
115. Warburg O. On the origin of cancer cells. *Science* 1956; 123 : 309-14.
116. Dang CV, Semenza GL. Oncogenic alterations of metabolism. *Trends Biochem Sci* 1999; 24 : 68-72.
117. Kim JW, Tchernyshyov I, Semenza GL, Dang CV. HIF-1-mediated expression of pyruvate dehydrogenase kinase: a metabolic switch required for cellular adaptation to hypoxia. *Cell Metab* 2006; 3 : 177-85.
118. Cuezva JM, Krajewska M, Heredia ML, Krajewski S, Kim GSH, Zapata JM, *et al*. The Bioenergetic signature of cancer: A marker of tumor progression. *Cancer Res* 2002; 62 : 6674-81.
119. Reivich M, Kuhl D, Wolf A, Greenberg J, Phelps M, Ido T, *et al*. Measurement of local cerebral glucose metabolism in man with 18F-2-fluoro-2-deoxy-d-glucose. *Acta Neurol Scand Suppl* 1977; 64 : 190-1.
120. Dichiro G, Brooks RA, Patronas NT, Bairamian D, Kornblith PL, Smith BH, *et al*. Issues in the *in vivo* measurement of glucose metabolism of human central nervous system tumours. *Ann Neurol* 1984; 15 : S138-46.
121. Padma MV, Said S, Jacobs M, Hwang DR, Dunigan K, Satter M, *et al*. Prediction of pathology and survival by FDG PET in gliomas. *J Neurooncol* 2003; 64 : 227-37.
122. Spence AM, Muzi M, Graham MM, O'Sullivan F, Link JM, Lewellen TK, *et al*. 2-[(18)F]Fluoro-2-deoxyglucose and glucose uptake in malignant gliomas before and after radiotherapy: correlation with outcome. *Clin Cancer Res* 2002; 8 : 971-9.
123. Weber WA. Positron emission tomography as an imaging biomarker. *J Clin Oncol* 2006; 20 : 3282-92.
124. Quennet V, Yaromina A, Zips D, Rosner A, Walenta S, Baumann M, *et al*. Tumor lactate content predicts for response to fractionated irradiation of human squamous cell carcinomas in nude mice. *Radiother Oncol* 2006; 81 : 130-5.
125. Gillies RJ, Robey I, Gatenby RA. Causes and consequences of increased glucose metabolism of cancers. *J Nuc Med* 2008; 49 : 24S-42S.
126. Mathupala SP, Ko YH, Pedersen PL. Hexokinase II: cancer's double-edged sword acting as both facilitator and gatekeeper of malignancy when bound to mitochondria. *Oncogene* 2006; 25 : 4777-86.
127. Dwarakanath BS, Zolzer F, Chandna S, Bauch T, Adhikari JS, Muller WU, *et al*. Heterogeneity in 2-deoxy-D-glucose induced modifications in energetic and radiation responses of human tumor cell lines. *Int J Radiat Oncol Biol Phys* 2001; 51 : 1151-61.
128. Xu RH, Pelicano H, Zhou Y, Carew JS, Feng L, Bhalla KN, *et al*. Inhibition of glycolysis in cancer cells: a novel strategy to overcome drug resistance associated with mitochondrial respiratory defect and hypoxia. *Cancer Res* 2005; 65 : 613-21.
129. Dwarakanath BS, Agrawala PK, Bhatt AN, Goenka S, Gupta D. Biological radioprotection: current status and prospects. *Radiat Protect Environ* 2008; 31 : 14-7.
130. Landau BR, Laszlo J, Stengle J, Burk D. Certain metabolic and pharmacologic effects in cancer patients given infusions of 2-deoxy-D-glucose. *J Natl Cancer Inst* 1958; 2 : 485-94.
131. Dwarakanath BS, Jain V. Targeting sweetness of tumors for improving cancer therapy *Future Oncol* 2009; 5 : 581-5.
132. Dwarakanath BS, Mathew TL, Jain V. Experimental and clinical studies to improve the radiotherapy of human brain tumors. *Biomedicine* 2000; 20 : 73-86.
133. Dwarakanath BS, Jain VK. Enhancement of radiation damage by 2-deoxy-D-glucose in organ cultures of brain tumours. *Indian J Med Res* 1985; 82 : 266-8.
134. Mohanti BK, Rath GK, Anantha N, Kannan V, Das BS, Chandramouli BA, *et al*. Improving cancer radiotherapy with 2-deoxy-D-glucose: Phase I/II clinical trials on human cerebral gliomas. *Int J Radiat Oncol Biol Phys* 1996; 35 : 103-11.
135. Singh D, Banerji AK, Dwarakanath BS, Tripathi RP, Gupta JP, Mathew TL, *et al*. Optimizing cancer radiotherapy with 2-deoxy-D-glucose: Dose escalation studies in patients with glioblastoma multiforme. *Strahlentherapie* 2005; 181 : 507-14.
136. Jain VK, Pohlit W, Purohit SC. Influence of energy metabolism on the repair of X-ray damage in living cells. III Effects of 2-deoxy-D-glucose on the liquid holding reactivation in yeast. *Biophysik* 1973; 10 : 137-42.
137. Jain VK, Kalia VK, Sharma R, Maharajan V, Menon M. Effects of 2-DG on glycolysis, proliferation kinetics and radiation response of human cancer cells. *Int J Radiat Oncol Biol Phys* 1985; 11 : 943-50.
138. Aft RL, Lewis JS, Zhang F, Kim J, Welch MJ. Enhancing targeted radiotherapy by copper(II)diacetyl- bis(N4-methylthiosemicarbazone) using 2-deoxy-D-glucose. *Cancer Res* 2003; 63 : 5496-504.

139. Jain V. Modifications of radiation responses by 2-deoxy-D-glucose in normal and cancer cells. *Indian J Nucl Med* 1996; *11* : 8-17.
140. Maher JC, Savaraj N, Priebe W, Liu H, Lampidis TJ. Differential sensitivity to 2-deoxy-D-glucose between two pancreatic cell lines correlates with GLUT-1 expression. *Pancreas* 2005; *30* : e34-9.
141. Dwarakanath BS, Khaitan D, Mathur R. Inhibitors of topoisomerases as anticancer drugs: Problems and prospects. *Indian J Exp Biol* 2004; *42* : 649-59.
142. Gridley DS, Nutter RL, Mantik DW, Slater JM. Hyperthermia and radiation *in vivo*: effect of 2-deoxy-D-glucose. *Int J Radiat Oncol Biol Phys* 1985; *11* : 567-74.
143. Hunter AJ, Blekkenhorst GH. The effects of 2-deoxyglucose and amino-oxyacetic acid on the radiation response of mammalian cells *in vitro*. *Int J Radiat Biol* 1998; *73* : 311-24.
144. Dwarakanath BS, Khaitan D, Ravindranath T. Two-deoxy-D-glucose enhances the cytotoxic effects of topoisomerase inhibitors in human tumor cell lines. *Cancer Biol Ther* 2004; *3* : 34-43.
145. Simons AL, Ahmad IM, Mattson DM, Dornfeld KJ, Spitz DR. 2-Deoxy-D-glucose combined with cisplatin enhances cytotoxicity via metabolic oxidative stress in human head and neck cancer cells. *Cancer Res* 2007; *67* : 3364-70.
146. Jain VK, Purohit SC, Pohlit W. Optimization of cancer therapy: Part I--Inhibition of repair of x-ray induced potentially lethal damage by 2-deoxy-D-glucose in Ehrlich-ascites tumour cells. *Indian J Exp Biol* 1977; *15* : 711-3.
147. Khaitan D, Chandna S, Arya MB, Dwarakanath BS. Differential mechanisms of radiosensitization by 2-deoxy-D-glucose in the monolayers and multicellular spheroids of a human glioma cell line. *Cancer Biol Ther* 2006; *5* : 1142-51.
148. Hay N, Sonenberg N. Upstream and downstream of mTOR. *Genes Dev* 2004; *18* : 1926-45.
149. Feng Z, Zhang H, Levine AJ, Jin S. The coordinate regulation of the p53 and mTOR pathways in cells. *Proc Natl Acad Sci USA* 2005; *102* : 8204-9.
150. Tee AR, Proud CG. Staurosporine inhibits phosphorylation of translational regulators linked to mTOR. *Cell Death Differ* 2001; *8* : 841-9.
151. Easton JB, Houghton PJ. Therapeutic potential of target of rapamycin inhibitors. *Expert Opin Ther Targets* 2005; *8* : 551-64.
152. Abraham RT, Gibbons JJ. The mammalian target of rapamycin signaling pathway: twists and turns in the road to cancer therapy. *Clin Cancer Res* 2007; *13* : 3109-14.
153. Blackburn EH. Telomere states and cell fates. *Nature* 2000; *408* : 53-6.
154. Forsyth NR, Wright WE, Shay JW. Telomerase and differentiation in multicellular organisms: Turn it off, turn it on, and turn it off again. *Differentiation* 2002; *69* : 188-97.
155. Shay JW, Bacchetti S. A survey of telomerase activity in human cancer. *Eur J Cancer* 1997; *5* : 787-91.
156. Shay JW, Wright WE. Telomerase therapeutics for cancer: challenges and new directions. *Nat Rev Drug Disc* 2006; *5* : 577-84.
157. Shay JW. Telomerase in cancer: diagnostic, prognostic, and therapeutic implications. *Cancer J Sci Am* 1998; (Suppl 1): S26-S34.
158. Li H, Liu JP. Signalling on telomerase: a master switch in cell aging and immortalization. *Biogerontology* 2002; *3* : 109-18.
159. Wong JMY, Kusdra L, Collins K. Subnuclear shuttling of human telomerase induced by transformation and DNA damage. *Nat Cell Biol* 2002; *4* : 731-6.
160. Herbert BS, Gellert GC, Hochreiter A, Pongracz K, Wright WE, Zielinska D, *et al*. Lipid modification of GRN163, an N3'→P5' thio-phosphoramidate oligonucleotide, enhances the potency of telomerase inhibition. *Oncogene* 2005; *24* : 5262-8.
161. Levine AJ, Finlay CA, Hinds PW. P53 is a tumor suppressor gene. *Cell* 2004; *116* : S67-9.
162. Lowe SW, Cepero E, Evan G. Intrinsic tumour suppression. *Nature* 2004; *432* : 307-15.
163. Erster S, Moll UM. Stress-induced p53 runs a direct mitochondrial death program: its role in physiologic and pathophysiological stress responses *in vivo*. *Cell Cycle* 2004; *3* : 1492-5.
164. Harms K, Nozell S, Chen X. The common and distinct target genes of the p53 family transcription factors. *Cell Mol Life Sci* 2004; *61* : 822-42.
165. Stoklosa T, Golab J. Prospects for p53-based cancer therapy. *Acta Biochim Pol* 2005; *52* : 321-8.
166. Lang FF, Bruner JM, Fuller GN, Aldape K, Prados MD, Chang S, *et al*. Phase I trial of adenovirus-mediated p53 gene therapy for recurrent glioma: biological and clinical results. *J Clin Oncol* 2003; *21* : 2508-18.
167. Zeimet AG, Marth C. Why did p53 gene therapy fail in ovarian cancer? *Lancet Oncol* 2003; *4* : 415-22.
168. Scott SL, Earle JD, Gumerlock PH. Functional p53 increases prostate cancer cell survival after exposure to fractionated doses of ionizing radiation. *Cancer Res* 2003; *63* : 7190-6.
169. Baselga J. Targeting tyrosine kinases in cancer: The second wave. *Science* 2006; *312* : 1175-8.
170. Paul MK, Mukhopadhyay AK. Tyrosine kinase - Role and significance in cancer. *Int J Med Sci* 2004; *1* : 101-15.
171. Violette M, Helene RF. Role of histone N-terminal tails and their acetylation in nucleosome dynamics. *Mol Cell Biol* 2000; *19* : 7230-7.
172. Munster PN, Troso S, Rosen N, Rifkind R, Marks PA, Richon VM. The histone deacetylase inhibitor suberoyanilide hydroxamic acid induces differentiation of human breast cancer cells. *Cancer Res* 2001; *61* : 8492-7.
173. Noriyuki T, Julian DC, Takashi K, Dorina G, Jonathan SW, Sadie W, *et al*. Histone deacetylase inhibitors have a profound antigrowth activity in endometrial cancer cells. *Cancer Res* 2004; *10* : 1141-9.
174. Coffey DC, Kutko MC, Glick RD, Butler LM, Heller G, Rifkind RA, *et al*. The histone deacetylase inhibitor, CBHA, inhibits growth of human neuroblastoma xenografts *in vivo*, alone and synergistically with *all-trans* retinoic acid. *Cancer Res* 2001; *61* : 3591-4.

175. Rasheed W, Bishton M, Johnstone RW, Prince HM. Histone deacetylase inhibitors in lymphoma and solid malignancies. *Expert Rev Anticancer Ther* 2008; 8 : 413-32.
176. Byrd JC, Marcucci G, Parthun MR, Xiao JJ, Klisovic RB, Moran M, *et al*. A phase I and pharmacodynamic study of depsipeptide (FK228) in chronic lymphocytic leukemia and acute myeloid leukemia. *Blood* 2005; 105 : 959-67.
177. Marchion DC, Bicaku E, Daud AI, Sullivan DM, Munster PN. *In vivo* synergy between topoisomerase II and histone deacetylase inhibitors: predictive correlates. *Mol Cancer Ther* 2005; 4 : 1993-2000.
178. Kristeleit R, Stimson L, Workman P, Aherne W. Histone modification enzymes: novel targets for cancer drugs. *Expert Opin Emerg Drugs* 2004; 9 : 135-54.
179. Lu KP, Suizu F, Zhou XZ, Finn G, Lam P, Wulf G. Targeting carcinogenesis: a role for the prolyl isomerase Pin1? *Mol Carcinog* 2006; 45 : 397-402.
180. Ryo A, Suizu F, Yoshida Y, Perrem K, Liou YC, Wulf G, *et al*. Regulation of NF-kappaB signaling by Pin1-catalyzed prolyl isomerization and ubiquitin-mediated proteolysis of p65/RelA. *Mol Cell* 2003; 12 : 1413-26.
181. Ryo A, Liou YC, Wulf G, Nakamura N, Lee SW, Lu KP. Pin1 is an E2F target gene essential for the Neu/Ras-induced transformation of mammary epithelial cells. *Mol Cell Biol* 2002; 22 : 5281-95.
182. Basu A, Das M, Qanungo S, Fan XJ, DuBois G, Haldar S. Proteasomal degradation of human peptidyl prolyl isomerase pin1-pointing phosphor Bcl2 toward dephosphorylation. *Neoplasia* 2002; 4 : 218-27.
183. Atchison FW, Capel B, Means AR. Pin1 regulates the timing of mammalian primordial germ cell proliferation. *Development* 2003; 130 : 3579-86.
184. Zheng H, You H, Zhou XZ, Murray SA, Uchida T, Wulf G, *et al*. The prolyl isomerase Pin1 is a regulator of p53 in genotoxic response. *Nature* 2002; 419 : 849-53.
185. Zacchi P, Gostissa M, Uchida T, Salvagno C, Avolio A, Voliniak S, *et al*. The prolyl isomerase Pin1 reveals a mechanism to control p53 functions after genotoxic insults. *Nature* 2002; 419 : 853-7.
186. Rippmann FJ, Hobbie S, Daiber C, Guilliard B, Bauer M, Birk J, *et al*. Phosphorylation-dependent proline isomerization catalyzed by Pin1 is essential for tumor cell survival and entry into mitosis. *Cell Growth Differ* 2000; 11 : 409-16.
187. Hennig L, Christner C, Kipping M, Scelbert B, Rucknagel KP, Grabley S, *et al*. Selective inactivation of parvulin like peptidyl-prolyl cis/trans isomerases by juglone. *Biochemistry* 1998; 37 : 5953-60.
188. Mathur R. *Experimental studies on the modification of cellular responses to topoisomerase inhibitors in normal and transformed cell lines*, Ph. D. thesis. University of Delhi; 2008.
189. Mathur R, Suman S, Beaume N, Ali M, Bhatt AN, Chopra M, *et al*. Interaction, structural modification of topoisomerase II α by peptidyl prolyl isomerase, Pin1: an *in silico* study. *Protein Pept Lett* 2010; 17 : 151-63.
190. Mathur R, Chandna S, Kapoor PN, Dwarakanath BS. Peptidyl prolyl isomerase, Pin1 is a potential target for enhancing the therapeutic efficacy of etoposide. *Curr Cancer Drug Targ* 2010; in press.

Reprint requests: Dr B.S. Dwarakanath, Division of Radiation Biosciences, Institute of Nuclear Medicine & Allied Sciences
Brig S.K. Mazumdar Road, Delhi 110 054, India
e-mail: bsd@inmas.drdo.in